

# A global accounting of medically significant scorpions: Epidemiology, major toxins, and comparative resources in harmless counterparts



Micaiah J. Ward\*, Schyler A. Ellsworth<sup>1</sup>, Gunnar S. Nystrom<sup>1</sup>

Department of Biological Science, Florida State University, Tallahassee, FL 32306, USA

## ARTICLE INFO

### Keywords:

Scorpion  
Venom  
Scorpionism  
Scorpion envenomation  
Scorpion distribution

## ABSTRACT

Scorpions are an ancient and diverse venomous lineage, with over 2200 currently recognized species. Only a small fraction of scorpion species are considered harmful to humans, but the often life-threatening symptoms caused by a single sting are significant enough to recognize scorpionism as a global health problem. The continued discovery and classification of new species has led to a steady increase in the number of both harmful and harmless scorpion species. The purpose of this review is to update the global record of medically significant scorpion species, assigning each to a recognized sting class based on reported symptoms, and provide the major toxin classes identified in their venoms. We also aim to shed light on the harmless species that, although not a threat to human health, should still be considered medically relevant for their potential in therapeutic development. Included in our review is discussion of the many contributing factors that may cause error in epidemiological estimations and in the determination of medically significant scorpion species, and we provide suggestions for future scorpion research that will aid in overcoming these errors.

## 1. Introduction

Originating approximately 450 million years ago, scorpions have since diversified into 19 recognized families and over 2200 species (Sharma et al., 2015; Lourenço, 2018). Epidemiological reviews on scorpionism have uncovered the dangerous reality of this global health problem, which results in thousands of deaths annually, and have contributed to the expansion of recognized harmful scorpion species (Müller, 1993; Al-Sadoon and Jarrar, 2003; Chippaux and Goyffon, 2008; Sari et al., 2011; Dehghani and Fathi, 2012; Borges et al., 2012; Santibáñez-López et al., 2015; Santos et al., 2016; Shahi et al., 2016; Bavani et al., 2017; Erickson and Cheema, 2017; Kang and Brooks, 2017; Riaño-Umbarila et al., 2017; Sanaei-Zadeh et al., 2017). Ten years ago, Chippaux and Goyffon (2008) listed 34 scorpion species known to cause human harm, with all but one (*Hemiscorpius lepturus*) belonging to the Buthidae family. It is now estimated that nearly 50 scorpion species are harmful to humans (Lourenço, 2018) and include the families Buthidae, Hemiscorpidae, and Scorpionidae. The majority of scorpion species, however, have not been reported in the literature as causing human harm and are generally considered harmless.

Scorpion venoms are a rich source of protein-based toxins, many of which have been identified as responsible for the painful and often life-threatening symptoms, especially the highly expressed ion-channel

toxins (Possani et al., 1999; de la Vega and Possani, 2004; de la Vega et al., 2010; Quintero-Hernández et al., 2013). Functional characterization of scorpion toxins has led to the development of life-saving medications, including a chlorotoxin found in *Leiurus hebraeus* (formerly *L. quinquestriatus hebraeus*), which can act as both an optical imaging contrast agent in the surgical removal of tumors, known as tumor paint, as well as an inhibitor of glioma cell invasion (Castle and Strong, 1986; Veiseh et al., 2007; Deshane et al., 2003). Scorpion venom characterization has also revealed that harmless scorpion species produce a plethora of toxins homologous to those found in their deadly relatives, including ion-channel toxins and antimicrobial peptides (Schwartz et al., 2007; Ma et al., 2009; Diego-García et al., 2012; He et al., 2013; Luna-Ramírez et al., 2015; Quintero-Hernández et al., 2015; Rokyta and Ward, 2017; Santibáñez-López et al., 2017; Ward et al., 2018). The term “medically significant” has been applied to scorpion species that cause human harm throughout the literature, often with the implication that venom from these species may be therapeutically useful. Harmless scorpion species, however, are just as medically relevant in drug development due to the homologous toxins ubiquitous in scorpion venoms.

The goal of this review is to provide an up-to-date global accounting of scorpion species identified as being medically significant in the literature, including geography, estimated sting frequency, symptoms,

\* Corresponding author. Florida State University, Department of Biological Science, 319 Stadium Dr., Tallahassee, FL 32306-4295 USA.

E-mail address: [mward@bio.fsu.edu](mailto:mward@bio.fsu.edu) (M.J. Ward).

<sup>1</sup> Contributed equally to manuscript.

and sting class assignments based on criteria proposed by Khattabi et al. (2011). Where available, the major toxin classes identified in their venoms are also reported to provide reference of scorpion toxin diversity and identify where additional venom characterization work is needed. We also highlight a few well-characterized scorpion species that are considered harmless to humans to illustrate their potential role in medicine and in understanding the evolutionary trajectories that have led to the co-existence of extraordinarily similar venoms with drastically different consequences. We recognize that several factors make an accurate assessment on this scale nearly impossible, including the continued emergence and reclassification of species, overlapping geographical regions of species with extremely similar morphology, variation in human sensitivity, and lack of proper scorpion identification. Nevertheless, we have made every effort to provide a summary of scorpions considered to be medically significant or harmful to humans as reported throughout the literature, with notes on contributing factors that may cause error in epidemiological estimations.

## 2. Methods

### 2.1. Search strategy

Our search strategy focused on scorpions that have been reported as medically significant or cause human harm in previous scorpionism literature reviews (Müller, 1993; Al-Sadoon and Jarrar, 2003; Chippaux and Goyffon, 2008; Sari et al., 2011; Borges et al., 2012; Dehghani and Fathi, 2012; Lourenço, 2015, 2018; Santibáñez-López et al., 2015; Santos et al., 2016; Shahi et al., 2016; Bavani et al., 2017; Erickson and Cheema, 2017; Kang and Brooks, 2017; Riaño-Umbarila et al., 2017; Sanaei-Zadeh et al., 2017; Salazar et al., 2018), and updating these records to include any species reported as harmful in the literature, or that required medical attention. Searches were performed in March–May of 2018 using traditional search tools such as PubMed and Google Scholar, as well as searching through literature available on the Virtual Health Library (VHL) following methods described by Santos et al. (2016), and reports publicly available from poison control centers (i.e. National Poison Data System annual reports). We did not limit searches to specific terms (i.e. “scorpion”, “public health”, etc.), as our goal was to find any available information, including epidemiology, geographic distribution, and venom characterization, on scorpion species that have been previously reported as medically significant. We do not include scorpion identification information as this has been discussed in many of the previously mentioned reviews and elsewhere (Lourenço, 2016; Rein, 2018), although the need for proper scorpion identification in epidemiological reporting is discussed. Due to the continued diversification of scorpion species and updated taxonomy classifications, we have retained species names as reported in the corresponding cited literature and noted taxonomic updates if available. Sting classifications, following criteria proposed by Khattabi et al. (2011), were only assigned to scorpions where symptoms attributed to that species were provided and verifiable. Sting frequencies were estimated based on the number of envenomations reported as being attributed to each species in the referenced literature and are meant to reflect the likely envenomation events for each species, although these are likely underestimations.

### 2.2. Map creation

All maps were generated using the ggplot2 package in R (Wickham, 2016). Mapped regions were scored as the number of species present in that location that have been reported as medically significant in the literature, such that locations with a greater number of medically significant species are darker than locations with fewer species. The maps do not reflect the number of envenomations or severity of symptoms by region.

### 2.3. Toxin abbreviations

Functional characterizations, descriptions, and definitions of toxin classes are outside the scope of this review and are discussed elsewhere, including Possani et al. (1999); de la Vega and Possani (2004); Zeng et al. (2005); de la Vega et al. (2010); Quintero-Hernández et al. (2013); Serrano (2013); Carmo et al. (2014); Harrison et al. (2014), as well as in many of the citations included throughout our review. We therefore only included major toxin classes that have been identified for each species by method of functional assay, individual toxin isolation, transcriptomic and/or proteomic approaches. Abbreviations are as follows: AMPs—antimicrobial peptides, Bpps—bradykinin-potentiating peptides, CaTxs—calcium-channel toxins, ClTxs—chloride-channel toxins, CRISPs—cysteine-rich secretory proteins, HYALs—hyaluronidases, KTxs—potassium-channel toxins, KUNs—Kunitz-type toxins, MPs—metalloproteases, NaTxs—sodium-channel toxins, PLA2s—phospholipases, SPs—serine proteases.

## 3. Results and discussion

Our search resulted in a total of 104 scorpion species considered medically significant or harmful to humans, including 101 Buthidae, 2 Hemiscorpidae, and 1 Scorpionidae. Sting classes were assigned following definitions described by Khattabi et al. (2011). Class I describes minor, localized symptoms that rarely require medical treatment. Class II describes moderate to severe symptoms that, although not life-threatening, usually do require medical treatment. **Class III describes severe, life-threatening symptoms that are likely fatal without medical treatment. All known scorpion species should be considered sting class I (harmless) unless otherwise documented.** Of the 104 scorpion species identified as medically significant in the literature, only 36 species were assigned a sting class of I–III based on symptoms reported as specific to that species. Four species were assigned to sting class I, eight species to sting class II, and 24 to sting class III. The remaining 68 species were given an unknown sting class because we were unable to verify symptoms for these species.

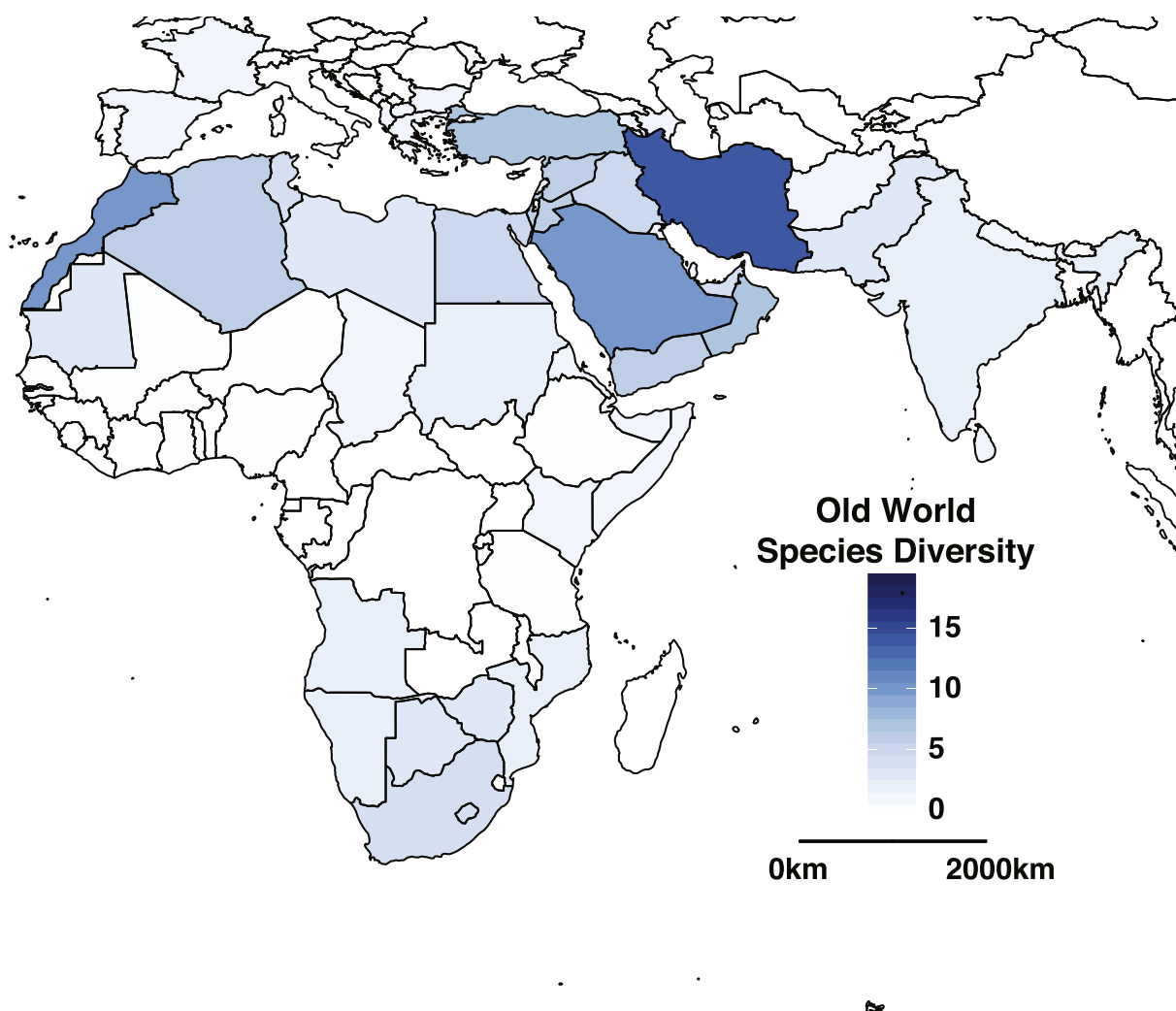
The global distributions of the medically significant species identified by our literature review are shown in Figs. 1 and 2. In the Old World (Fig. 1), the darker locations corresponding to Iran, Saudi Arabia, and Morocco, indicate the density of medically significant scorpion species found in these areas, with fewer found in the surrounding countries of Africa, Asia, and Europe (Table 1). In the New World (Fig. 2), it appears that Mexico, Brazil, and Venezuela, are rich with harmful scorpion fauna, with fewer harmful species described from the United States, Central America, the Caribbean, and other South American countries (Table 2).

Venom characterization can include the isolation and functional characterization of individual toxins, venom-gland transcriptome sequencing, and/or venom proteomics, all of which contribute to and are necessary for the development of therapeutics from venom. Fifty-four of the 104 scorpion species we identified as medically significant had some type of venom characterization work completed, many of which were limited to the isolation of one or a handful of toxins from the venom. Only 12 species had undergone more extensive venom characterizations including transcriptomic or proteomic approaches. We could not find venom characterization studies for the remaining 51 medically significant scorpion species.

### 3.1. Family Buthidae

#### 3.1.1. *Apistobuthus*

We found one species belonging to the *Apistobuthus* genus, *A. pterygocercus*, which is distributed throughout Middle Eastern countries of Asia (Table 1). Al-Sadoon and Jarrar (2003) referenced this species as medically significant in Saudi Arabia, however, no specific case reports were found that could associate specific envenomation symptoms with



**Fig. 1. Old World heat map of medically significant scorpion diversity.** Higher scores (darker regions) indicate countries with a greater number of reported medically significant species, and lower scores (lighter regions) indicate countries with fewer reported medically significant species (see methods). The map does not reflect the number of envenomations or severity of symptoms by region.

this species. Dehghani and Fathi (2012) also mentioned *A. pterygocercus* as being medically significant, but with only minor importance. For these reasons, we included *A. pterygocercus* in our review with an unknown sting classification, as it is unclear whether or not this species is harmful to humans (Hauke and Herzig, 2017). We could not find any confirmed venom toxins for *A. pterygocercus*.

### 3.1.2. *Androctonus*

We found six species belonging to the *Androctonus* genus, *A. amoreuxi*, *A. australis*, *A. bicolor* (*A. aeneas*), *A. crassicauda*, *A. liouvillei*, and *A. mauretanicus*. Members of *Androctonus* are commonly referred to as “fat-tailed” scorpions, and are distributed in parts of Africa and Asia (Table 1). Recent taxonomy of *Androctonus* (Coelho et al., 2014) and epidemiological reports (Chakroun-Walha et al., 2018) indicate *A. bicolor* and *A. aeneas* as being the same species and are counted as such for the purposes of this review, although others have indicated that further taxonomic clarification is needed (Goyffon et al., 2012). *Androctonus amoreuxi*, *A. bicolor* (*A. aeneas*), and *A. liouvillei*, have been reported as harmful to humans, however, these reports are from areas with high scorpionism rates and no symptoms were assigned to specific species (Touloun et al., 2001; Coelho et al., 2016; Amr et al., 2017). Additionally, *A. amoreuxi* has previously been reported as harmless (Goyffon et al., 2012). We therefore included *A. amoreuxi*, *A. bicolor* (*A. aeneas*), and *A. liouvillei*, in our list with an unknown sting classification

(Table 3). *Androctonus australis*, *A. crassicauda*, and *A. mauretanicus* have been responsible for severe envenomation symptoms, including fatalities, and were assigned to sting class III (Table 3).

Partial venom characterizations have been completed for *A. australis* (Laraba-Djebari et al., 1994), *A. crassicauda* (Caliskan et al., 2013), and *A. mauretanicus* (Martin-Eauclaire and Bougis, 2012), all of which revealed potent NaTxS and KTxs that contribute to potentially fatal stings. A complete venom characterization using transcriptomic and proteomic analyses was completed for *A. bicolor* (*A. aeneas*), which identified NaTxS, KTxs, CaTxS, AMPs, defensins, KUNs, and Bpps (Zhang et al., 2015). Another group isolated two AMPs from the venom of *A. bicolor* (*A. aeneas*), and synthetic analogues of these peptides showed potent antimicrobial and anticancer activities (Du et al., 2015). Several toxins have also been isolated from *A. amoreuxi* including AMPs (Almaaytah et al., 2012) and ion-channel toxins (Chen et al., 2003, 2005), and this venom has been shown to be effective against multiple cancer lines (Salem et al., 2016; Akef et al., 2017). We could not find any confirmed toxins for *A. liouvillei*.

### 3.1.3. *Buthacus*

We identified one member of the *Buthacus* genus, *B. macrocentrus*, which is distributed throughout Middle Eastern countries of Asia (Table 1). We could not find any specific case reports or envenomation symptoms reported for this species (Table 3), however, Caliskan et al.

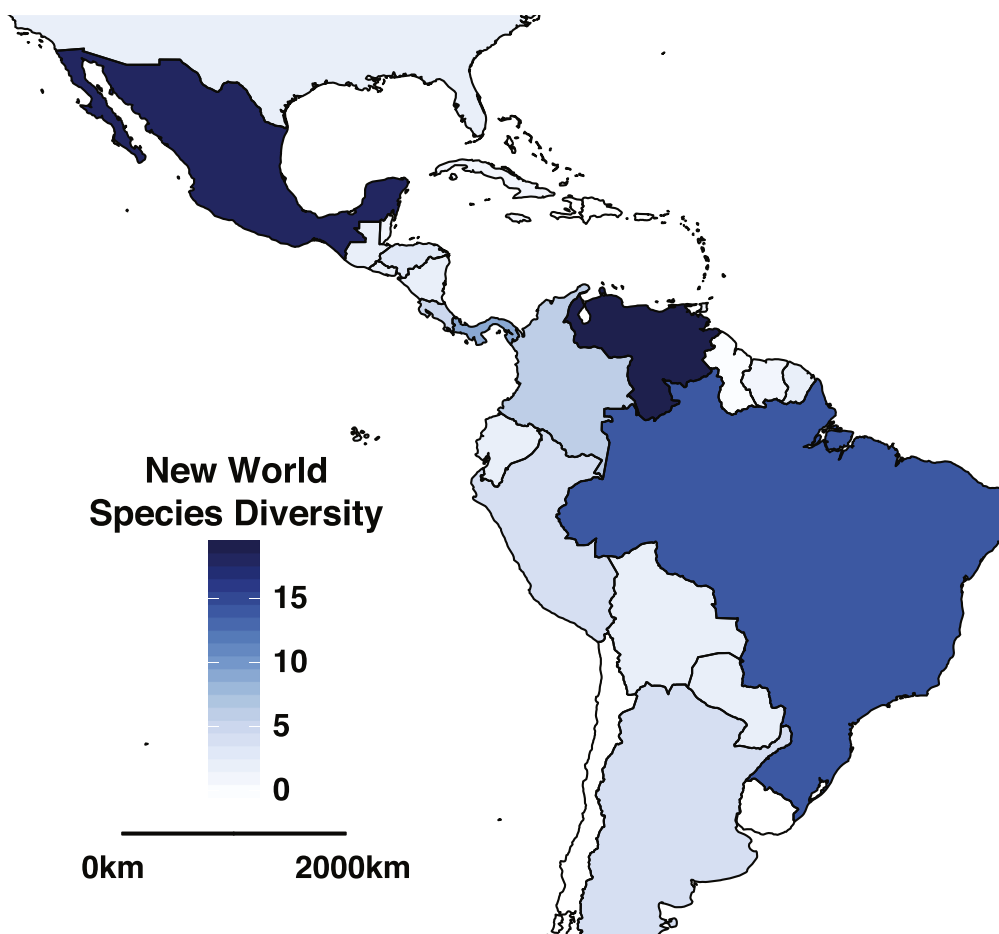


Fig. 2. New World heat map of medically significant scorpion diversity. Higher scores (darker regions) indicate countries with a greater number of reported medically significant species, and lower scores (lighter regions) indicate countries with fewer reported medically significant species (see methods). The map does not reflect the number of envenomations or severity of symptoms by region.

(2012) suggested this species be characterized as dangerous to humans based on the likelihood of being misidentified as either a *Leiurus* or *Mesobuthus* species (these are all described as “yellow” scorpions in Turkey), the functional characterization of a NaTx in the venom, and the high toxicity of *B. macrocentrus* venom in mice. Although the lethality in one vertebrate is not necessarily an indicator of lethality in another, even when toxins are deemed mammal-specific (van der Meijden et al., 2017), we still included *B. macrocentrus* in our review with an unknown sting classification (Table 3) as it is unclear whether or not this species should be considered medically significant.

#### 3.1.4. *Buthus*

In our initial searches, we only found one medically significant *Buthus* species, *B. occitanus*, which was widely distributed throughout the Old World (Fet et al., 2000). Recent taxonomical updates have reclassified several new species that were formerly considered *B. occitanus*, and there are currently 52 species of *Buthus* distributed throughout Europe, Africa, and Asia, with *B. occitanus* having a much more limited range (Sousa et al., 2017). Previous reports attributed severe envenomations to *B. occitanus* (Ghalim et al., 2000; Aboumaâd et al., 2014), however, these reports were based in Morocco where it is unlikely that *B. occitanus* exists. According to Sousa et al. (2017), *B. occitanus* is limited to France and Spain, and there are 17 other *Buthus* species in Morocco. El Hidan et al. (2017) lists 12 *Buthus* species in Morocco, including *B. occitanus*, and Emerich et al. (2017) performed a comparative analysis on four subspecies of *B. occitanus* from Morocco that Sousa et al. (2017) classified as full species. Because the taxonomical updates to *Buthus* by Sousa et al. (2017) were unavailable at

the time El Hidan et al. (2017) and Emerich et al. (2017) were published, we elected to follow the classification and location information provided by Sousa et al. (2017) for the purposes of this review.

The recent taxonomical updates for this genus have not yet been widely incorporated into the literature and the species of medically significant *Buthus* are not yet clear. We conducted separate searches for each of the 52 *Buthus* species listed by Sousa et al. (2017), and although we did not find case reports or symptoms assigned to a specific species, we did find reports suggesting at least six *Buthus* species should be considered medically significant and have included them in our list with an unknown sting classification (Table 3). *Buthus tunetanus*, *B. paris*, *B. malhommei*, and *B. mardochei*, were all toxic at low doses when injected into mice, and all have KTx and NaTx present in their venoms (Emerich et al., 2017). Hmimou et al. (2008) also reported *B. malhommei* as being responsible for severe envenomations in humans in Morocco, although the report did not separate the number of scorpion envenomations caused by *B. malhommei* from other scorpions in the area. *Buthus lienhardi* (Laaradia et al., 2018) and *B. occitanus* (Martin-Eauclaire et al., 2014) venoms were also shown to be toxic at low doses when injected into mice.

NaTx were identified as the most abundant and diverse group of toxins in the venom of *B. occitanus* from France, where the species is government protected (Martin-Eauclaire et al., 2014). In a study using *Buthus* from Egypt (formerly *B. occitanus*, now five separate *Buthus* species), Bpps were identified as one of the most abundant toxins (Meki et al., 1995).

**Table 1**  
Old World geographic distribution of medically significant scorpions.

Species	Region	Country(ies)	Source(s)
<b>Family Buthidae</b>			
<b>Apistobuthus</b>			
<i>A. pterygocercus</i>	Asia	Iran, Kuwait, Oman, Qatar, Saudi Arabia, United Arab Emirates, Yemen	Dehghani and Fathi (2012)
<b>Androctonus</b>			
<i>A. amoreuxi</i>	Africa	Egypt, Mali, Morocco	Goyffon et al. (2012); Coelho et al. (2014); Amr et al. (2017); Badry et al. (2018); El Hidan et al. (2018)
<i>A. australis</i>	Asia	Jordan	Chippaux and Goyffon (2008); Coelho et al. (2014); Amr et al. (2017); Badry et al. (2018)
	Africa	Algeria, Chad, Egypt, Libya, Mauritania, Morocco, Somalia, Sudan, Tunisia	Fet et al. (2000); Coelho et al. (2014); Amr et al. (2017); Badry et al. (2018)
<i>A. bicolor</i> ( <i>A. aeneas</i> )	Asia	India, Israel, Jordan, Pakistan, Saudi Arabia, Yemen	Fet et al. (2000); Coelho et al. (2014); Amr et al. (2017); Badry et al. (2018)
	Africa	Algeria, Egypt, Eritrea, Libya, Morocco, Tunisia	Fet et al. (2000); Coelho et al. (2014); Amr et al. (2017); Badry et al. (2018)
<i>A. crassicauda</i>	Asia	Israel, Jordan, Lebanon, Syria	Fet et al. (2000); Amr et al. (2017); Badry et al. (2018)
	Africa	Egypt, Mauritania, Morocco	Amr et al. (2017); Badry et al. (2018)
<i>A. liouvillei</i>	Asia	Armenia, Azerbaijan, Bahrain, Iran, Iraq, Israel, Jordan, Kuwait, Oman, Saudi Arabia, Syria, Turkey, United Arab Emirates, Yemen	Coelho et al. (2014); El Hidan et al. (2017)
	Africa	Morocco	Fet et al. (2000); Touloun et al. (2012); Coelho et al. (2014); El Hidan et al. (2017)
<i>A. mauretanicus</i>	Africa	Algeria, Mauritania, Morocco	
<b>Buthacus</b>			
<i>B. macrocentrus</i>	Asia	Bahrain, Iran, Iraq, Israel, Jordan, Oman, Qatar, Saudi Arabia, Syria, Turkey, United Arab Emirates	Yağmur et al. (2008)
<b>Buthus</b>			
<i>B. lienhardi</i>	Africa	Morocco	Sousa et al. (2017)
<i>B. malhommei</i>	Africa	Morocco	Sousa et al. (2017)
<i>B. mardochei</i>	Africa	Morocco	Sousa et al. (2017)
<i>B. occitanus</i>	Europe	France, Spain	Sousa et al. (2017)
<i>B. paris</i>	Africa	Algeria, Morocco, Tunisia	Sousa et al. (2017)
<i>B. tunetanus</i>	Africa	Algeria, Lybia, Morocco, Tunisia	Sousa et al. (2017)
<b>Compsobuthus</b>			
<i>C. matthiesseni</i>	Asia	Iran	Navidpour (2015)
<i>C. persicus</i>	Asia	Iran	Navidpour (2015)
<b>Hottentotta</b>			
<i>H. gentili</i>	Africa	Algeria, Morocco	Kovařík (2007)
<i>H. jayakari</i>	Asia	Iran, Oman, Saudi Arabia, United Arab Emirates, Yemen	Sari et al. (2011); Kovařík (2007)
<i>H. saulcyi</i>	Asia	Afghanistan, Iran, Iraq, Turkey	Kovařík (2007)
<i>H. schach</i>	Asia	Iran, Iraq	Kovařík (2007)
<i>H. tamulus</i>	Asia	India, Nepal, Pakistan, Sri Lanka	Kovařík (2007); Strong et al. (2015)
<i>H. zagrosensis</i>	Asia	Iran	Kovařík (2007); Sanaei-Zadeh et al. (2017)
<b>Leiurus</b>			
<i>L. abdullahbayrami</i>	Asia	Syria and Turkey	Khalil and Yağmur (2010); Lowe et al. (2014)
<i>L. arabicus</i>	Asia	Saudi Arabia	Lowe et al. (2014)
<i>L. brachycentrus</i>	Asia	Saudi Arabia, Yemen	Lowe et al. (2014)
<i>L. haenggii</i>	Asia	Oman, Saudi Arabia, Yemen	Lowe et al. (2014)
<i>L. heberti</i>	Asia	Oman	Lowe et al. (2014)
<i>L. hebraeus</i>	Asia	Israel, Jordan, Lebanon, Syria	Lowe et al. (2014); Amr et al. (2017)
<i>L. jordanensis</i>	Asia	Jordan, Saudi Arabia	Lowe et al. (2014); Amr et al. (2017)
<i>L. macroctenus</i>	Asia	Oman	Lowe et al. (2014)
<i>L. quinquestriatus</i>	Asia	Egypt, Sudan	Lowe et al. (2014); Badry et al. (2018)
<b>Mesobuthus</b>			
<i>M. caucasicus</i> complex	Asia	Iran, Turkey	Fet et al. (2018)
<i>M. eupeus</i>	Asia	Iran, Turkey	Karataş and Karataş (2003); Ozkan and Kat (2005); Dehghani and Fathi (2012); Navidpour (2015)
<i>M. gibbosus</i>	Asia	Turkey	Ozkan and Kat (2005); Parmakelis et al. (2006); Kaltsas et al. (2008)
	Europe	Albania, Bulgaria, Greece, Macedonia (FYROM), Montenegro	
<b>Odontobuthus</b>			
<i>O. doriae</i>	Asia	Iran	Lourenco and Pezier (2002)
<b>Orthochirus</b>			
<i>O. scrobiculosus</i>	Asia	Iran	Dehghani and Fathi (2012)
<b>Parabuthus</b>			
<i>P. granulatus</i>	Africa	Angola, Botswana, Kenya, Namibia, South Africa, Zimbabwe	Fet et al. (2000); Prendini and Esposito (2010)
<i>P. mossambicensis</i>	Africa	Botswana, Mozambique, South Africa, Zimbabwe	Fet et al. (2000)
<i>P. transvaalicus</i>	Africa	Botswana, Mozambique, South Africa, Zimbabwe	Fet et al. (2000)
<i>P. villosus</i>	Africa	Angola, Namibia, South Africa	Fet et al. (2000); Prendini and Esposito (2010)
<b>Family Hemiscorpidae</b>			
<b>Hemiscorpius</b>			
<i>H. acanthocercus</i>	Asia	Iran	Navidpour (2015)
<i>H. lepturus</i>	Asia	Iran, Iraq, Oman, Pakistan, Saudi Arabia, United Arab Emirates, Yemen	Navidpour (2015); Dehghani et al. (2018)
<b>Family Scorpionidae</b>			
<b>Nebo</b>			
<i>N. hierichonticus</i>	Africa	Egypt	Rosin and Shulov (1963);
	Asia	Israel, Jordan, Saudi Arabia, Syria	Badry et al. (2018)

### 3.1.5. *Centruroides*

The *Centruroides* genus is one of the most diverse scorpion genera, currently comprised of 90 species (Esposito et al., 2017). Several

*Centruroides* species are commonly referred to as bark scorpions as they are often arboreal and found under peeling tree bark (Jiménez-Jiménez and Palacios-Cardiel, 2010; Esposito et al., 2017). The majority of

**Table 2**  
New World geographic distribution of medically significant scorpions.

Species	Region	Country(ies)	Source(s)
<b>Family Buthidae</b>			
<b>Centruroides</b>			
<i>C. balsasensis</i>	North America	Mexico	Ponce Saavedra and Francke (2004); Riaño-Umbarila et al. (2017)
<i>C. bicolor</i>	Central America	Panama	Salazar et al. (2018)
<i>C. elegans</i>	North America	Mexico	Riaño-Umbarila et al. (2017)
<i>C. exilicauda</i>	North America	Mexico	Jiménez-Jiménez and Palacios-Cardiel (2010)
<i>C. gracilis</i>	Caribbean	Cuba, Jamaica	Sissom and Lourenço (1987);
	Central America	Belize, El Salvador, Guatemala, Honduras	Fet et al. (2000); Teruel (2008);
	North America	Mexico, United States	Borges et al. (2012)
	South America	Northern regions	Otero et al. (2004); Borges et al. (2012)
<i>C. granosus</i>	Central America	Panama	Salazar et al. (2018)
<i>C. hirsutipalpus</i>	North America	Mexico	Riaño-Umbarila et al. (2017)
<i>C. huichol</i>	North America	Mexico	Teruel et al. (2015)
<i>C. infamatus</i>	North America	Mexico	Riaño-Umbarila et al. (2017)
<i>C. limbatus</i>	Central America	Costa Rica, Honduras, Nicaragua, Panama	Borges et al. (2012); Salazar et al. (2018)
<i>C. limpidus</i>	North America	Mexico	Riaño-Umbarila et al. (2017)
<i>C. margaritatus</i>	Central America	Costa Rica, El Salvador, Guatemala, Honduras, Nicaragua, Panama	Sissom and Lourenço (1987); Fet et al. (2000);
	North America	Mexico	Borges et al. (2012)
	South America	Western Colombia	
<i>C. meisei</i>	North America	Mexico	Riaño-Umbarila et al. (2017)
<i>C. noxius</i>	North America	Mexico	Riaño-Umbarila et al. (2017)
<i>C. ornatus</i>	North America	Mexico	Riaño-Umbarila et al. (2017)
<i>C. panamensis</i>	Central America	Panama	Salazar et al. (2018)
<i>C. pococki</i>	Caribbean	Lesser Antilles	Fet et al. (2000)
<i>C. sculpturatus</i>	North America	United States, Mexico	Riaño-Umbarila et al. (2017); Kang and Brooks (2017)
<i>C. suffusus</i>	North America	Mexico	Riaño-Umbarila et al. (2017)
<i>C. tecomanus</i>	North America	Mexico	Riaño-Umbarila et al. (2017)
<i>C. testaceus</i>	Caribbean	Lesser Antilles	Fet et al. (2000)
	South America	Venezuela	
<i>C. villegasi</i>	North America	Mexico	Riaño-Umbarila et al. (2017)
<i>C. sp. nov A</i>	North America	Mexico	Riaño-Umbarila et al. (2017)
<i>C. sp. nov B</i>	North America	Mexico	Riaño-Umbarila et al. (2017)
<b>Tityus</b>			
<i>T. apiacas</i>	South America	Brazil	Silva et al. (2017)
<i>T. arellanoparrai</i>	South America	Venezuela	D'Suze et al. (2015)
<i>T. asthenes</i>	Central America	Costa Rica, Ecuador, Panama	Fet et al. (2000); Otero et al. (2004); Salazar et al. (2018)
	South America	Brazil, Colombia, Peru	
<i>T. bahiensis</i>	South America	Argentina, Brazil, Paraguay	Brasil (2009); Lourenço (2015)
<i>T. bastosi</i>	Central America	Ecuador	Fet et al. (2000)
	South America	Brazil, Colombia, Peru	Costa et al. (2016)
<i>T. breweri</i>	South America	Venezuela	Fet et al. (2000); Borges et al. (2010); D'Suze et al. (2015)
<i>T. carabobensis</i>	South America	Venezuela	D'Suze et al. (2015)
<i>T. caripitensis</i>	South America	Venezuela	D'Suze et al. (2015)
<i>T. cerroazul</i>	Central America	Costa Rica, Panama	Borges et al. (2012); Salazar et al. (2018)
<i>T. confuatus</i>	South America	Argentina, Bolivia, Brazil, Paraguay	Lourenço and da Silva (2007); Brasil (2009)
<i>T. costatus</i>	South America	Brazil	Fet et al. (2000); Brasil (2009); Lourenço (2015)
<i>T. discrepans</i>	South America	North-Central Venezuela	Borges et al. (2011)
<i>T. falconensis</i>	South America	Venezuela	D'Suze et al. (2015)
<i>T. festae</i>	Central America	Panama	Borges et al. (2012);
	South America	Colombia	Salazar et al. (2018)
<i>T. feuhrmanni</i>	South America	Colombia	Otero et al. (2004)
<i>T. isabelceciliae</i>	South America	Venezuela	Borges et al. (2010); D'Suze et al. (2015)
<i>T. ivic-nancor</i>	South America	Venezuela	D'Suze et al. (2015)
<i>T. matthieseni</i>	South America	Brazil	Costa et al. (2016)
<i>T. metuendus</i>	South America	Brazil, Peru	Fet et al. (2000); Lourenço (2008); Brasil (2009)
<i>T. monaguensis</i>	South America	Venezuela	D'Suze et al. (2015)
<i>T. neoespartanus</i>	South America	Venezuela	Fet et al. (2000); De Sousa et al. (2007)
<i>T. nororientalis</i>	South America	Venezuela	D'Suze et al. (2015)
<i>T. obscurus</i>	South America	Brazil, French Guiana, Suriname	Fet et al. (2000); Brasil (2009); Pardal et al. (2014); Torrez et al. (2015)
<i>T. pachyurus</i>	Central America	Costa Rica, Panama	Fet et al. (2000); Otero et al. (2004); Salazar et al. (2018)
	South America	Colombia, Venezuela	
<i>T. perijanensis</i>	South America	Venezuela	Fet et al. (2000); D'Suze et al. (2015)
<i>T. pittieri</i>	South America	Venezuela	D'Suze et al. (2015)
<i>T. pusillus</i>	South America	Brazil	Dias et al. (2006); Porto et al. (2010)
<i>T. quirogae</i>	South America	Venezuela	D'Suze et al. (2015)
<i>T. sanarensis</i>	South America	Venezuela	D'Suze et al. (2015)
<i>T. serrulatus</i>	South America	Argentina, Bolivia, Brazil	Fet et al. (2000); Brasil (2009); Lourenço (2015)
<i>T. silvestris</i>	South America	Brazil, French Guiana, Peru	Fet et al. (2000); Brasil (2009); Monteiro et al. (2016)
<i>T. stigmurus</i>	South America	Brazil	Brasil (2009); Bertani et al. (2018)
<i>T. surorientalis</i>	South America	Venezuela	D'Suze et al. (2015)
<i>T. trinitatis</i>	South America	Trinidad and Tobago	Daisley et al. (1999)
<i>T. trivittatus</i>	South America	Argentina, Brazil	de Roodt (2014)
<i>T. valarae</i>	South America	Venezuela	D'Suze et al. (2015)
<i>T. zulianus</i>	South America	Venezuela	Borges et al. (2011)

**Table 3**  
**Reported envenomations and symptoms of medically significant scorpions.** Annual sting frequencies were estimated based on cited literature if available, and are likely still underestimates. A sting class was assigned if symptoms reported, based on the Khattabi et al. (2011) classification system. In brief: Class I indicates minor, localized symptoms; Class II indicates non life-threatening, moderate to severe systemic symptoms; Class III indicates severe systemic symptoms that are life-threatening if untreated. Those with a U indicate the sting class is unknown.

Species	Estimated Annual Sting Frequency	Source(s)	Sting Class	Reported Symptoms	Source(s)
<b>Family Buthidae</b>					
<b>Apistobuthus</b>					
<i>A. pterygocercus</i>	100–1000	Al-Sadoon and Jarrar (2003)	U	unknown, no specific case reports or symptoms found, reported as being of minor importance	Dehghani and Fathi (2012)
<b>Androctonus</b>					
<i>A. amoreuxi</i>	Unknown		U	No specific case reports or symptoms found. Reported as both harmful and harmless in literature	Goyffon et al. (2012); Coelho et al. (2014); Amr et al. (2017)
<i>A. australis</i>	100–1000	Chakroun-Walaha et al. (2018)	III	tachycardia, tachypnea, heart failure, pulmonary edema	Bahloul et al. (2004, 2005)
<i>A. bicolor (A. aeneus)</i>	10–100	Chakroun-Walaha et al. (2018)	U	unknown, may be fatal but no specific case reports or symptoms found	Touloun et al. (2001); Coelho et al. (2016)
<i>A. crassicauda</i>	1000–10,000	Dehghani et al. (2009); Ozkan et al. (2006); Al-Sadoon and Jarrar (2003)	III	pain and burning at sting site, tachypnea, drowsiness, gastrointestinal pain, diarrhea, vomiting, increased salivation, numbness, hypotension, hypothermia, cyanosis, can be fatal within 9–36 h	Radmanesh (1990); Dehghani and Fathi (2012)
<i>A. liouvillei</i>	Unknown		U	reported as medically significant, formerly <i>A. bicolor liouvillei</i> , no specific case reports or symptoms found	Coelho et al. (2014)
<i>A. mauretanicus</i>	100–1000	Ghalim et al. (2000); Touloun et al. (2012)	III	pain and burning sensation, sweating, shivering, hyperthermia, severe cases can be fatal	Ghalim et al. (2000); Touloun et al. (2001)
<b>Buthacus</b>					
<i>B. macrocentrus</i>	Unknown		U	unknown, no specific case reports or symptoms found, may have been misidentified as <i>Leiurus</i> or <i>Mesobuthus</i> species	Caliskan et al. (2012)
<b>Buthus</b>					
<i>B. lienhardi</i>	10–100	Ghalim et al. (2000); Aboumaad et al. (2014)	U	unknown, formerly <i>B. occitanus</i>	Ghalim et al. (2000); Aboumaad et al. (2014); Sousa et al. (2017)
<i>B. malhommei</i>	10–100	Ghalim et al. (2000); Hmimou et al. (2008); Aboumaad et al. (2014)	U	unknown, formerly <i>B. occitanus</i>	Ghalim et al. (2000); Hmimou et al. (2008); Aboumaad et al. (2014); Sousa et al. (2017)
<i>B. mardochei</i>	10–100	Ghalim et al. (2000); Aboumaad et al. (2014)	U	unknown, formerly <i>B. occitanus</i>	Ghalim et al. (2000); Aboumaad et al. (2014); Sousa et al. (2017)
<i>B. occitanus</i>	< 10	Martin-Eauclaire et al. (2014)	U	unknown, no specific case reports or symptoms found after species reclassification	Martin-Eauclaire et al. (2014); Sousa et al. (2017)
<i>B. paris</i>	10–100	Ghalim et al. (2000); Aboumaad et al. (2014)	U	unknown, formerly <i>B. occitanus</i>	Ghalim et al. (2000); Aboumaad et al. (2014); Sousa et al. (2017)
<i>B. tunetanus</i>	10–100	Ghalim et al. (2000); Aboumaad et al. (2014)	U	unknown, formerly <i>B. occitanus</i>	Ghalim et al. (2000); Aboumaad et al. (2014); Sousa et al. (2017)
<b>Compsothatus</b>					
<i>C. mathieseni</i>	10–100	Dehghani et al. (2009); Sanaei-Zadeh et al. (2017); Shahi et al. (2016)	U	unknown, aside from requiring medical attention and some reports of hematuria that require confirmation	Dehghani et al. (2009); Sanaei-Zadeh et al. (2017); Shahi et al. (2016)
<i>C. persicus</i>	10–100	Shahi et al. (2016)	U	unknown, aside from requiring medical attention	Shahi et al. (2016)
<b>Centruroides</b>					
<i>C. balsasensis</i>	100–1000	Riño-Umbarila et al. (2017)	I	pain, numbness	Ponce Saavedra and Francke (2004)
<i>C. bicolor</i>	100–1000	Salazar et al. (2018)	U	reported as causing severe envenomations, case reports need confirmation	Salazar et al. (2018)
<i>C. elegans</i>	100–1000	Riño-Umbarila et al. (2017)	U	described as deadly, with no specific case reports or symptoms found	Riño-Umbarila et al. (2017); Vandendriessche et al. (2010)
<i>C. exilicauda</i>	10–100	Chippaux and Goyffon (2008)	U	unknown, previously reported as harmful before differentiating from <i>C. sculpturatus</i>	Valdez-Gruz et al. (2004); Chippaux and Goyffon (2008)
<i>C. gracilis</i>	10–100	Otero et al. (2004)	II	pain, systemic symptoms, may be region-dependent	Otero et al. (2004); Borges et al. (2012)
<i>C. granosus</i>	100–1000	Salazar et al. (2018)	U	unknown, reported as dangerous, no specific case reports or symptoms found	Salazar et al. (2018)
<i>C. hirsutipalpus</i>	100–1000	Riño-Umbarila et al. (2017)	U	no specific case reports or symptoms found	Riño-Umbarila et al. (2017)
<i>C. huichal</i>	Unknown		U	unknown, formerly <i>C. noxius</i> before classified as separate species	(Teruel et al., 2015)
<i>C. infamatus</i>	100–1000	Riño-Umbarila et al. (2017)	U	no specific case reports or symptoms found	Riño-Umbarila et al. (2017)
<i>C. limbatus</i>	100–1000	Bush (1999); Salazar et al. (2018)	II	pain, paresthesia, flushing, hypertension, wheezing	Bush (1999)
<i>C. limpidus</i>	100–1000	Riño-Umbarila et al. (2017)	U	no specific case reports or symptoms found	Riño-Umbarila et al. (2017)

(continued on next page)

Table 3 (continued)

Species	Estimated Annual Sting Frequency	Source(s)	Sting Class	Reported Symptoms	Source(s)
<i>C. margaritatus</i>	10–100	Marinkelle and Stahnke (1965); Borges et al. (2012)	I	pain, local edema, fever, mild symptoms	Marinkelle and Stahnke (1965); Borges et al. (2012)
<i>C. mensei</i>	100–1000	Riño-Umbarila et al. (2017)	U	no specific case reports or symptoms found	Riño-Umbarila et al. (2017)
<i>C. noxius</i>	100–1000	Riño-Umbarila et al. (2017)	U	described as the most toxic species in Mexico, responsible for adult human deaths, but no specific case reports or symptoms found	Teruel et al. (2015); Riño-Umbarila et al. (2017)
<i>C. ornatus</i>	100–1000	Riño-Umbarila et al. (2017)	U	no specific case reports or symptoms found	Riño-Umbarila et al. (2017)
<i>C. panamensis</i>	100–1000	Salazar et al. (2018)	U	unknown, reported as dangerous, no specific case reports or symptoms found	Salazar et al. (2018)
<i>C. pococki</i>	< 10	Schmitt et al. (2017)	I	pain at sting site, numbness in lips and fingers	Schmitt et al. (2017)
<i>C. sculpuratus</i>	10–100	Chippaux and Goyffon (2008); Mowry et al. (2015, 2016); Gummin et al. (2017); Riño-Umbarila et al. (2017)	III	pain at sting site, tachycardia, restlessness, roving eye movements, hypertension, respiratory distress, tachypnea, hypersalivation, slurred speech, stridor, can be fatal	Skolnik and Ewald (2013); Kang and Brooks (2017)
<i>C. suffusus</i>	100–1000	Riño-Umbarila et al. (2017)	U	no specific case reports or symptoms found	Riño-Umbarila et al. (2017)
<i>C. tecomanus</i>	100–1000	Riño-Umbarila et al. (2017)	U	described as lethal, but no specific case reports or symptoms found	Valdez-Velázquez et al. (2013); Olamendi-Portugal et al. (2016); Riño-Umbarila et al. (2017)
<i>C. tesracus</i>	< 10	Lobo et al. (2011)	I	pain at sting site, erythema	Lobo et al. (2011)
<i>C. villegasi</i>	100–1000	Riño-Umbarila et al. (2017)	U	no specific case reports or symptoms found	Riño-Umbarila et al. (2017)
<i>C. sp. nov. A</i>	100–1000	Riño-Umbarila et al. (2017)	U	no specific case reports or symptoms found	Riño-Umbarila et al. (2017)
<i>C. sp. nov. B</i>	100–1000	Riño-Umbarila et al. (2017)	U	no specific case reports or symptoms found	Riño-Umbarila et al. (2017)
<b>Hottentotta</b>					
<i>H. gentili</i>	10–100	Touloun et al. (2012)	III	severe envenomations requiring medical treatment reported, including 10 fatalities	Touloun et al. (2012)
<i>H. jayakari</i>	< 10	Sanaei-Zadeh et al. (2017); Suresh et al. (2014)	II	infection of bone marrow reported	Suresh et al. (2014)
<i>H. saulyi</i>	< 10	Dehghani et al. (2009); Shahi et al. (2016)	U	unknown, although required medical treatment	Dehghani et al. (2009); Shahi et al. (2016)
<i>H. schach</i>	< 10	Dehghani et al. (2009)	U	unknown, although required medical treatment	Dehghani et al. (2009)
<i>H. tamulus</i>	10–100	Bawaskar and Bawaskar (1998); Kularatne et al. (2015)	III	pain and numbness at sting site, swelling, tachycardia, increased blood pressure, sweating, salivation, hypotension, hypertension, piloerection and priapism, vomiting, parasternal systolic lift, cardiac arrhythmias, cold extremities, pulmonary edema, heart failure	Bawaskar and Bawaskar (1998); Kularatne et al. (2015)
<i>H. zagrosensis</i>	< 10	Sanaei-Zadeh et al. (2017)	U	unknown, although required medical treatment	Sanaei-Zadeh et al. (2017)
<b>Leiurus</b>					
<i>L. abdullahbayrami</i>	< 10	Seiter et al. (2016); Dokur et al. (2017)	III	pain, burning and blistering at sting site, tachycardia, nausea, sweating, hallucinations, hypotension, cardiomyopathy, pulmonary edema	Seiter et al. (2016); Dokur et al. (2017)
<i>L. arabicus</i>	100–1000	Al-Sadoon and Jarrar (2003)	U	unknown, formerly <i>L. quinquestriatus</i>	Al-Sadoon and Jarrar (2003); Lowe et al. (2014)
<i>L. brachycentrus</i>	100–1000	Al-Sadoon and Jarrar (2003)	U	unknown, formerly <i>L. quinquestriatus</i>	Al-Sadoon and Jarrar (2003); Lowe et al. (2014)
<i>L. haenggi</i>	100–1000	Al-Sadoon and Jarrar (2003)	U	unknown, formerly <i>L. quinquestriatus</i>	Al-Sadoon and Jarrar (2003); Lowe et al. (2014)
<i>L. heberti</i>	10–100	Al-Sadoon and Jarrar (2003)	U	unknown, formerly <i>L. quinquestriatus</i>	Al-Sadoon and Jarrar (2003); Lowe et al. (2014)
<i>L. hebraeus</i>	100–1000	Al-Sadoon and Jarrar (2003); Amr et al. (2017)	III	formerly <i>L. quinquestriatus</i> , lethargy, confusion, pulmonary edema, cardiogenic shock, severe hypertension, pancreatitis, can be fatal	Sofer and Gueron (1988); Sofer et al. (1991); Lowe et al. (2014)
<i>L. jordanensis</i>	100–1000	Al-Sadoon and Jarrar (2003); Amr et al. (2017)	U	unknown, formerly <i>L. quinquestriatus</i>	Al-Sadoon and Jarrar (2003); Lowe et al. (2014)
<i>L. macrotenus</i>	10–100	Al-Sadoon and Jarrar (2003)	U	unknown, formerly <i>L. quinquestriatus</i>	Al-Sadoon and Jarrar (2003); Lowe et al. (2014)
<i>L. quinquestriatus</i>	10–100	Saad et al. (2017)	U	unknown, recently reclassified into several new species	Lowe et al. (2014)
<b>Mesobuthus</b>					
<i>M. caucasicus</i> complex	Unknown	Ozkan and Kat (2005); Chippaux and Goyffon (2008); Cesaretti and Ozkan (2010)	U	unknown, recently classified into several subspecies (see section 3.1.9)	Ozkan and Kat (2005)
<i>M. eupeus</i>	100–1000		III	pain, swelling, and burning at sting site, dry mouth, thirst, sweating, hypotension, hyperemia	Ozkan and Kat (2005)

(continued on next page)

Table 3 (continued)

Species	Estimated Annual Sting Frequency	Source(s)	Sting Class	Reported Symptoms	Source(s)
<i>M. gibbosus</i>	10–100	Chippaux and Goyffon (2008); Cesaretli and Ozkan (2010)	U	systemic manifestations and fatalities reported, case reports need confirmation	Ozkan and Ciftci (2010)
<b><i>Odontobutius</i></b>					
<i>O. doriae</i>	< 10	Razi and Malekanrad (2008); Dehghani and Fathi (2012)	III	pain and swelling at sting site, disorientation, erythema, acute asymmetric pulmonary edema, tachypnea, tachycardia	Razi and Malekanrad (2008); Dehghani and Fathi (2012)
<b><i>Orthochirus</i></b>					
<i>O. scrobiculosus</i>	< 10	(Dehghani and Fathi, 2012)	U	unknown, reported as causing fatality as well as minor symptoms, case reports need confirmation	(Dehghani et al., 2009; Hauke and Herzig, 2017)
<b><i>Parabuthus</i></b>					
<i>P. granularis</i>	10–100	Müller (1993); Bergman (1997)	III	pain and burning at sting site, difficulty swallowing, muscle pain and cramps, restlessness, potential respiratory failure, can be fatal	Müller (1993)
<i>P. mossambicensis</i>	10–100	Bergman (1997)	U	unknown, likely mild to moderate envenomation symptoms	Bergman (1997)
<i>P. transvaalicus</i>	100–1000	Bergman (1997)	III	pain and swelling at sting site, difficulty swallowing, muscle tremors or myoclonic jerks, fasciculation of the tongue, hypersalivation, sweating, bilateral ptosis, difficulty passing urine, can be fatal	Bergman (1997)
<i>P. villosus</i>	Unknown		U	unknown, likely mild envenomation symptoms, considered medically important in early literature but no other envenomation reports found	Debont et al. (1998)
<b><i>Tityus</i></b>					
<i>T. apicatus</i>	< 10	Silva et al. (2017)	II	pain and redness at sting site, systemic manifestations	Silva et al. (2017)
<i>T. arellanoparrai</i>	< 10	D'Suza et al. (2015)	U	described as causing severe and fatal accidents, but no specific case reports or symptoms found	D'Suza et al. (2015)
<i>T. asthenes</i>	100–1000	Otero et al. (2004); Borges et al. (2012); Salazar et al. (2018)	III	pain, burning and redness at sting site, edema, persistent vomiting, cerebral decortication, lack of corneal reflex, distended abdomen, peripheral cyanosis, cardio-respiratory failure, tachypnea, hypoxaemia, hyperamylasaemia, pancreatitis, can be fatal	Otero et al. (2004); Borges et al. (2015)
<i>T. bahiensis</i>	10–100	Bucarechchi et al. (2014)	III	cardiovascular manifestations, respiratory failure, can be fatal	de Oliveira et al. (2015)
<i>T. basiosus</i>	10–100	Costa et al. (2016)	U	unknown, although required medical treatment	Costa et al. (2016)
<i>T. breweri</i>	< 10	Borges et al. (2010); D'Suza et al. (2015)	II	pain and redness at sting site, sweating, hypersalivation, muscle tremors, fever, piloerection, tachycardia, tachypnea, abdominal pain, described as causing fatal accidents (needs confirmation)	Borges et al. (2010); D'Suza et al. (2015)
<i>T. carabobensis</i>	< 10	D'Suza et al. (2015)	U	described as causing severe and fatal accidents, but no specific case reports or symptoms found	D'Suza et al. (2015)
<i>T. caripitensis</i>	< 10	D'Suza et al. (2015)	U	described as causing severe and fatal accidents, but no specific case reports or symptoms found	D'Suza et al. (2015)
<i>T. cerroazul</i>	100–1000	Borges et al. (2012); Salazar et al. (2018)	U	fatalities reported, case reports need confirmation	Borges et al. (2012)
<i>T. confluentis</i>	< 10	de Roodt et al. (2009)	III	can be fatal	de Roodt et al. (2009)
<i>T. costatus</i>	10–100	Diego-García et al. (2005)	U	unknown, other than symptoms require medical attention	Diego-García et al. (2005)
<i>T. discrepans</i>	10–100	D'suza et al. (2003); Borges et al. (2011)	III	hypertension, hypotension, tachycardia, tachypnea, hypothermia, respiratory distress, pancreatitis, gastrointestinal disorders, can be fatal	D'suza et al. (2003); Borges et al. (2011)
<i>T. falconensis</i>	< 10	D'Suza et al. (2015)	U	described as causing severe and fatal accidents, but no specific case reports or symptoms found	D'Suza et al. (2015)
<i>T. festae</i>	100–1000	Borges et al. (2012); Salazar et al. (2018)	U	fatalities reported, case reports need confirmation	Borges et al. (2012)
<i>T. fuehrmanni</i>	10–100	Otero et al. (2004)	U	unknown, described as moderate to severe requiring medical attention, but no specific symptoms reported	Otero et al. (2004)
<i>T. isabeleceitiae</i>	< 10	D'Suza et al. (2015)	U	described as causing severe and fatal accidents, but no specific case reports or symptoms found	D'Suza et al. (2015)
<i>T. ivic-nancor</i>	< 10	D'Suza et al. (2015)	U	described as causing severe and fatal accidents, but no specific case reports or symptoms found	D'Suza et al. (2015)
<i>T. mathieseni</i>	10–100	Costa et al. (2016)	U	unknown, although required medical treatment	Costa et al. (2016)
<i>T. metuendus</i>	< 10	Lourenço (2016)	U	fatalities reported, case reports need confirmation	Lourenço (2016)
<i>T. monaguensis</i>	< 10	D'Suza et al. (2015)	U	described as causing severe and fatal accidents, but no specific case reports or symptoms found	D'Suza et al. (2015)

(continued on next page)

Table 3 (continued)

Species	Estimated Annual Sting Frequency	Source(s)	Sting Class	Reported Symptoms	Source(s)
<i>T. neoespartanus</i>	< 10	De Sousa et al. (2007)	II	vomiting, labored breathing, muscle tremors, hypersalivation, sweating, piloerection, tachypnea, cardiac arrhythmia, edematous pancreatitis, leukocytosis, hyperglycemia, hyperamyloasemia	De Sousa et al. (2007)
<i>T. nororientalis</i>	< 10	D'Suze et al. (2015)	U	described as being responsible for severe and fatal accidents, no specific record or symptoms	D'Suze et al. (2015)
<i>T. obscurus</i>	10–100	Torrez et al. (2015); Lourenço (2016)	III	pain, burning and redness at sting site, edema, sweating, piloerection and pruritus, nausea, dizziness, blurred vision, tremors, agitation, electric shock-like sensations in the body, slurred speech, lack of coordination, can be fatal	Pardal et al. (2014)
<i>T. pachyurus</i>	100–1000	Otero et al. (2004); Izquierdo and Buitrago (2012); Salazar et al. (2018)	III	severe systemic symptoms causing myocardial dysfunction, cardiovascular collapse, heart arrest, respiratory failure, pulmonary edema	Izquierdo and Buitrago (2012)
<i>T. perijanensis</i>	< 10	D'Suze et al. (2015)	U	described as causing severe and fatal accidents, possible neurological manifestations, but no specific case reports	D'Suze et al. (2015)
<i>T. pititieri</i>	< 10	D'Suze et al. (2015)	U	described as causing severe and fatal accidents, but no specific case reports or symptoms found	D'Suze et al. (2015)
<i>T. pusillus</i>	< 10	Albuquerque et al. (2009)	II	pain and burning at sting site, chills, dizziness, headache and vomiting	Albuquerque et al. (2009)
<i>T. quitogae</i>	< 10	D'Suze et al. (2015)	U	described as causing severe and fatal accidents, but no specific case reports or symptoms found	D'Suze et al. (2015)
<i>T. sanarensis</i>	< 10	D'Suze et al. (2015)	U	described as causing severe and fatal accidents, but no specific case reports or symptoms found	D'Suze et al. (2015)
<i>T. serrulatus</i>	10–100	Bucaretschi et al. (2014)	III	pain at the site of the sting, hypersalivation, vomiting, sweating, psychomotor agitation, cardiac arrhythmias, arterial hypertension, pulmonary edema, circulatory failure	Teixeira et al. (2001); Cupo and Hering (2002); Bucaretschi et al. (2014)
<i>T. silvestris</i>	< 10	Coelho et al. (2016); Monteiro et al. (2016)	II	pain and burning at sting site, labored breathing, agitation, tachycardia, generalized muscle spasms, hypertension	Monteiro et al. (2016)
<i>T. stigmurus</i>	100–1000	Batista et al. (2007)	III	pain, burning and swelling at sting site, numbness, headache, vomiting, sweating, can be fatal	Lira-da-Silva et al. (2000); Batista et al. (2007)
<i>T. surorientalis</i>	< 10	D'Suze et al. (2015)	U	described as causing severe and fatal accidents, but no specific case reports or symptoms found	D'Suze et al. (2015)
<i>T. trinitatis</i>	< 10	Daisley et al. (1999)	III	tachypnea, restlessness, vomiting, hypersalivation, cerebral edema, pulmonary edema, hypovolemic shock, convulsions, myocarditis, pancreatitis	Daisley et al. (1999)
<i>T. trivittatus</i>	100–1000	de Roodt et al. (2003); de Roodt (2014)	III	pain, swelling, redness and burning at sting site, hyperthermia, cardiovascular and circulatory symptoms, cramps, tremors, headache, vomiting, dizziness, sweating, joint pain, can be fatal	de Roodt et al. (2003)
<i>T. valerae</i>	< 10	D'Suze et al. (2015)	U	described as causing severe and fatal accidents, but no specific case reports or symptoms found	D'Suze et al. (2015)
<i>T. zuluianus</i>	10–100	Borges et al. (2011)	U	local manifestations, respiratory arrest, pulmonary edema, and fatalities reported, case reports need confirmation	Chowell et al. (2006); Borges et al. (2011)
<b>Family Hemiscorpiidae</b>					
<b><i>Hemiscorpius</i></b>					
<i>H. acanthocercus</i>	100–1000	Chippaux and Goyffon (2008); Shahi et al. (2016); Dehghani et al. (2018)	III	excess proteins and blood in urine, hemolysis of blood cells, severe effects on blood and kidneys, can be fatal	Shahi et al. (2015); Dehghani et al. (2018)
<i>H. lepturus</i>	100–1000	Jalali et al. (2010); Chippaux and Goyffon (2008); Dehghani et al. (2018)	III	toxic action on blood cells, kidney and liver function, necrosis, can be fatal	Pipelzadeh et al. (2007); Jalali et al. (2010); Dehghani and Fathi (2012); Dehghani et al. (2018)
<b>Family Scorpionidae</b>					
<b><i>Nebo</i></b>					
<i>N. hierichonticus</i>	< 10	Annobil (1993)	III	pain, itching and swelling at sting site, intravascular coagulopathy, intracranial hemorrhage, pulmonary edema, congestive heart failure, can be fatal	Rosin and Shulov (1963); Annobil (1993)

*Centruroides* considered medically significant are located in Mexico, with a small number of species found in the Southern United States, South and Central America, and throughout the Caribbean (Table 2). We found 24 *Centruroides* species that were considered medically significant in the literature, although only seven were able to be assigned a sting classification (Table 3). Recently, Riaño-Umbarila et al. (2017) performed a comprehensive median lethal dose assay, using venoms from 14 *Centruroides* species from Mexico, including two new species yet to be formally described, all of which were toxic in mice. Although toxicity in mice should not be used to indicate medical significance to humans (van der Meijden et al., 2017), Riaño-Umbarila et al. (2017) also reported the number of scorpion envenomations by region and which scorpions were responsible based on a 2016 report by the Ministry of Health of the Mexican Government. For these reasons, we included the 14 *Centruroides* species studied by Riaño-Umbarila et al. (2017) in our list of medically significant species, and assigned a sting class to those with documented case studies and symptoms (Table 3). Salazar et al. (2018) performed a similar study on *Centruroides* species in Panama, with over 33,000 scorpion stings including 47 deaths documented by the Department of Epidemiology of the Ministry of Health between 2000 and 2016, which were attributed as being caused by a handful of *Centruroides* or *Tityus* species. Venom from the four *Centruroides* species included in the Salazar et al. (2018) study were reported as dangerous and shown to be toxic in mice, although only one, *C. limbatus*, was assigned a sting class of II based on confirmed severe envenomation symptoms (Table 3).

In addition to the more comprehensive studies of *Centruroides* from Mexico and Panama discussed above, we found six other *Centruroides* species identified as being medically significant in the literature. *Centruroides exilicauda*, formerly synonymous with *C. sculpturatus*, was identified as a separate species that appears to be less harmful compared to *C. sculpturatus* (Valdez-Cruz et al., 2004). *Centruroides gracilis* has documented moderate to severe envenomations in Colombia (Otero et al., 2004), although this species has been reported as mild in Belize (Borges et al., 2012). *Centruroides margaritatus*, which is distributed throughout Central and South America, appears to be associated with either mild or more severe envenomations depending on the region (Marinkelle and Stahnke, 1965; Borges et al., 2012). *Centruroides pococki* (Schmitt et al., 2017), and *C. testaceus* (Lobo et al., 2011) envenomations have been described as having mild to moderate symptoms. *Centruroides noxius* is considered the most toxic species in Mexico (Santibáñez-López et al., 2015; Riaño-Umbarila et al., 2017), however, we were unable to find a single case report or symptoms specifically associated with this species, and were therefore only able to assign *C. noxius* to an unknown sting class (Table 3).

Individual toxins isolated from *C. elegans* (Vandendriessche et al., 2010; Santibáñez-López et al., 2015; Olamendi-Portugal et al., 2005; Restano-Cassulini et al., 2008), *C. exilicauda* (Valdez-Cruz et al., 2004), *C. gracilis* (Possani et al., 2000), *C. infamatus* (Dehesa-Dávila et al., 1996), *C. limbatus* (Koschak et al., 1998), *C. limpidus* (Lebreton et al., 1994; Dehesa-Dávila et al., 1996; Santibáñez-López et al., 2016), *C. margaritatus* (García-Calvo et al., 1993), *C. sculpturatus* (Wang and Strichartz, 1983; Corona et al., 2001; Valdez-Cruz et al., 2004), and *C. suffusus* (Martin et al., 1987; Estrada et al., 2007; Corzo et al., 2008), revealed a rich diversity of NaTxS and KTxs with both insect and mammal specificity. NaTxS were also detected in the electrophoretic profiles of *C. hirsutipalpus*, *C. meisei*, *C. ornatus*, *C. villegasi*, *C. sp. nov. A*, and *C. sp. nov. B* (Riaño-Umbarila et al., 2017).

A limited number of *Centruroides* venoms have been more well-characterized using venom-gland transcriptomic and proteomic approaches. *Centruroides noxius* venom has been characterized using high-throughput transcriptomic and proteomic methods (Rendón-Anaya et al., 2012) and isolation of individual toxins (Possani et al., 1982; Zamudio et al., 1992), which have confirmed NaTxS and KTxs, as well as AMPs, MPs, and protease inhibitors. A partial venom-gland transcriptome and proteomic characterization of *C. tecomanus* also

identified NaTxS, KTxs, AMPs, and MPs (Valdez-Velazquez et al., 2013, 2016), some of which have been individually isolated and functionally characterized (Ramírez et al., 1988; Olamendi-Portugal et al., 2016).

### 3.1.6. *Compsobuthus*

We found two species belonging to the *Compsobuthus* genus, *C. matthiesseni* and *C. persicus*, both of which are present in Iran (Table 1). *Compsobuthus matthiesseni* was identified in several epidemiological and biodiversity reports (Dehghani et al., 2009; Dehghani and Fathi, 2012; Nejati et al., 2014; Navidpour, 2015; Sanaei-Zadeh et al., 2017) and was found to be responsible for over 20% of confirmed scorpion stings in Iran (Dehghani et al., 2009), although the severity of symptoms was not reported beyond requiring medical attention (Table 3). More recent studies have reported symptoms such as hematuria (Dehghani and Fathi, 2012), but the degree of harm this species has on humans is still unclear. *Compsobuthus persicus* was responsible for nearly 20% of scorpion stings in the south of Iran from 2014 to 2016 (Shahi et al., 2016), with no symptoms reported as being specific to this species. Both *C. matthiesseni* and *C. persicus* were assigned to an unknown sting class (Table 3), and we could not find any confirmed venom components for either species.

### 3.1.7. *Hottentotta*

We found six species belonging to the *Hottentotta* genus, *H. gentili*, *H. jayakari*, *H. saulcyi*, *H. schach*, *H. tamulus* (formerly *Mesobuthus tamulus*), and *H. zagrosensis*, all of which are found in parts of Asia, aside from *H. gentili*, which is found in Northern Africa (Table 1). Envenomations by *H. gentili* were assigned to sting class III due to the manifestation of life-threatening symptoms and at least 10 fatalities (Table 3). Very few *H. jayakari* envenomations have been reported (Table 3), but those that have, have required extensive medical treatment, which assigns *H. jayakari* to sting class II. We found documented envenomation reports for both *H. saulcyi* and *H. schach*, although these did not include symptoms specific to these species other than requiring medical treatment (Table 3). Of the five *Hottentotta* species identified, *H. tamulus* has the highest number of documented envenomations, including multiple deaths (Bawaskar and Bawaskar, 1998; Kularatne et al., 2015), and was assigned to sting class III (Table 3). Venom composition and symptoms caused by *H. tamulus* seem to vary depending on habitat and geography, where the coastal populations tend to cause more harm than those found in semi-arid inland plateaus (Strong et al., 2015). We found one confirmed case report of *H. zagrosensis* with no specific symptoms reported (Table 3).

We could not find any confirmed venom components for *H. jayakari* or *H. saulcyi*. In a partial venom characterization of *H. gentili* venom, SPs and PLA2s were confirmed and functionally characterized (Estrada-Gómez et al., 2017). NaTxS, KTxs, ClTxS, PLA2s, and pulmonary edema causing toxins, have all been confirmed in the venom of *H. tamulus* (Strong et al., 2001, 2015; Deshpande et al., 2005). We also found a study that functionally characterized NaTxS in the venom of *Buthotus schach* (Aboutorabi et al., 2016), which was said to be synonymized with *H. zagrosensis* in this reference. However, Kovařík (2007) synonymized *B. schach* with *H. schach* (not *H. zagrosensis*) and classified *H. schach* and *H. zagrosensis* as separate species. We have retained the position that *H. schach* and *H. zagrosensis* are separate species for the purposes of this review, and it is unclear which species was used in the study by Aboutorabi et al. (2016).

### 3.1.8. *Leiurus*

The *Leiurus* genus, once considered monospecific under *L. quinquestriatus*, has undergone extensive taxonomic reclassification over the last several years and is now comprised of 12 species, each with fairly distinct geographical ranges in North Africa and the Levant and Arabian Peninsula (Table 1). *Leiurus quinquestriatus* is the only *Leiurus* found in Egypt, where epidemiological studies have included *L. quinquestriatus* as one of the most probable culprits of severe scorpion envenomations

(Saad et al., 2017). However, other medically significant species are also found in Egypt and Saad et al. (2017) did not include specimen identification. Venom from *L. quinquestriatus* was shown to be toxic when injected into rats (Salman and Hammad, 2017), but the degree of severity in humans is difficult to assess without confirmed case reports. Based on the classifications by Lowe et al. (2014), many of the previous case reports attributed to *L. quinquestriatus* should now be reclassified. Sofer and Gueron (1988) and Sofer et al. (1991) case reports are from a specific hospital in Israel (Soroka Medical Center) which is located in the distribution range of *L. hebraeus*, making this species the most probable culprit of these envenomations (Table 3). Al-Sadoon and Jarrar (2003) also identified *L. quinquestriatus* as one of the three offending scorpions in Saudi Arabia, and based on the regions represented in their report, stings attributed to *L. quinquestriatus* should now be shared among *L. arabicus*, *L. brachycentrus*, *L. haenggii*, *L. hebraeus*, *L. jordanensis*, *L. macroctenus*, and *L. heberti* (Table 3). More recent reports cite *L. abdullahbayrami* as being medically significant and responsible for severe, but rare, envenomations (Seiter et al., 2016; Dokur et al., 2017). Amr et al. (2017) lists *L. hebraeus* and *L. jordanensis* as being two of the five harmful species in Jordan, with *L. hebraeus* being the most common, but could not specify the identity of scorpions responsible for the 1205 stings reported in Jordan between 2006 and 2012. Based on these reports, nine *Leiurus* species are included in our list of medically significant scorpion species, although only *L. abdullahbayrami* and *L. hebraeus* were assigned a sting classification (Table 3). We could not find any envenomation records or notes of medical significance for the three remaining African *Leiurus* species (Lourenço et al., 2006, 2018; Lourenço and Rossi, 2016), and these should be further investigated.

The reclassification of *Leiurus* species also has implications in medical development. The *Leiurus* chlorotoxin used as an optical imaging contrast agent, in the surgical removal of tumors (tumor paint), and as an inhibitor of glioma cell invasion, was originally identified by Castle and Strong (1986), who listed the venom source as being from *L. quinquestriatus hebraeus*, which is now *L. hebraeus*. Following the isolation of *Leiurus* chlorotoxin, the exact species from which the toxin was isolated was not always specified (Deshane et al., 2003; Veiseh et al., 2007), and some refer to isolation from an Israeli scorpion (Dardevet et al., 2015), which also implies *L. hebraeus*.

Peptidomic and functional characterization of *L. abdullahbayrami* venom suggests the presence of NaTx, ClTx, and AMPs (Dokur et al., 2017). In *L. hebraeus*, KTx and ClTx have been purified and functionally characterized (Castle and Strong, 1986; Garcia et al., 1994; Kumar et al., 2015). We did not find venom characterization information for any of the other *Leiurus* species.

### 3.1.9. *Mesobuthus*

We found three species belonging to the *Mesobuthus* genus, *M. caucasicus*, *M. eupeus* and *M. gibbosus*. *Mesobuthus caucasicus* has recently been reclassified into several species (Fet et al., 2018) which are sympatric with *M. eupeus* and *M. gibbosus* in Iran and Turkey (Table 1). Although we did not find specific envenomation reports or venom characterization for members of the *M. caucasicus* complex, some have suggested that these species may be harmful to humans and that comparative lethality assays and venom characterization should be completed (Adiguzel, 2010; Ozkan et al., 2008; Bavani et al., 2017). Both *M. eupeus* and *M. gibbosus* are likely responsible for hundreds of envenomations annually with severe envenomation symptoms reported (Table 3), although case reports for *M. gibbosus* need confirmation.

KTx and antimalarial peptides have been identified in *M. eupeus* (Gao et al., 2010a,b; Kuzmenkov et al., 2015), and KTx, ClTx, and AMPs, have been identified in *M. gibbosus* (Diego-García et al., 2013, 2014).

### 3.1.10. *Odontobuthus*

We found one species belonging to the *Odontobuthus* genus, *O. doriae*, which is a burrowing species distributed in Iran (Table 1).

Although we could only find one case report for *O. doriae*, it was included as a medically significant species causing moderate to severe envenomation symptoms by others (Dehghani and Fathi, 2012; Sanaei-Zadeh et al., 2017). Based on reported symptoms, *O. doriae* was assigned to a sting class of III (Table 3).

KTx and NaTx have been isolated and functionally characterized from the venom of *O. doriae* (Jalali et al., 2005; Abdel-Mottaleb et al., 2006), and the venom-gland transcriptome revealed the presence of KTx, NaTx, AMPs, and MPs (NaderiSoorki et al., 2016).

### 3.1.11. *Orthochirus*

We found one species belonging to the genus *Orthochirus*, *O. scrobiculosus*, which is distributed throughout Iran (Table 1). This species is described as docile with very low envenomation risk, even when directly handled (Dehghani and Fathi, 2012) and has been classified as not likely to cause severe human harm (Hauke and Herzig, 2017). This species has been reported to cause at least one fatality, with other symptoms being minor, although envenomation reports were unable to be verified (Dehghani and Fathi, 2012). We therefore included *O. scrobiculosus* in our review with an unknown sting classification (Table 3).

KTx have been isolated and functionally characterized from the venom of *O. scrobiculosus* (Dudina et al., 2001), and were shown to be lethal when injected into mice (Mouhat et al., 2005).

### 3.1.12. *Parabuthus*

We found four species belonging to the *Parabuthus* genus, *P. granulatus*, *P. mossambicensis*, *P. transvaalicus*, and *P. villosus*, all of which are distributed in central and southern countries of Africa (Table 1). *Parabuthus villosus* was identified as medically important by Debont et al. (1998), although we could not find any verified case reports for this species (Table 3). Envenomation symptoms caused by *P. mossambicensis* could also not be verified, but are likely mild to moderate (Table 3). Severe envenomation symptoms have been caused by *P. granulatus* and *P. transvaalicus*, assigning them to sting class III (Table 3). Prendini and Esposito (2010) updated the taxonomy of *Parabuthus* in South Africa and Namibia and found that several other *Parabuthus* species are sympatric with *P. granulatus*, which may indicate that some envenomation reports for *P. granulatus* could be misclassified. We could not find reports suggesting other *Parabuthus* species should be considered medically significant. NaTx and KTx have been identified in the venom from all four of the included *Parabuthus* species (Debont et al., 1998; Dyason et al., 2002).

### 3.1.13. *Tityus*

*Tityus* is the most diverse genus in Buthidae with over 200 described species widely distributed throughout Central and South America as well as in the Caribbean (Ojanguren-Affilastro et al., 2017). Unlike many of the other species described here, most *Tityus* species are accustomed to habitats with higher levels of moisture and humidity, such as tropical, coastal and rainforest habitats (Lourenço, 2008; Diego-García et al., 2012; Monteiro et al., 2016). Although we only list species that we found to be medically significant in the literature, our results are likely an underestimation due to the vast diversity and distribution of *Tityus*, with new species continuing to be classified (Lourenço, 2017; Teruel and de los Santos, 2018). We found 37 *Tityus* species of medical significance, 14 of which we were able to assign a sting class of either II or III based on reported symptoms (Table 3). Among those with unknown sting classifications were those described as causing severe or fatal accidents, but we could not find specific case reports to provide confirmation. In some cases, the severity of symptoms caused by members of the same species is dependent on the region. For example, *T. obscurus*, which is a senior synonym for both *T. paraensis* and *T. cambridgei* (Lourenço and Leguin, 2008), is found in both eastern and western regions of Parà, Brazil, with notable differences in venom composition and envenomation symptoms between these populations

(Pardal et al., 2014; Torrez et al., 2015; Santos et al., 2016). Both populations of *T. obscurus* can cause severe symptoms that require medical treatment, but the western population causes more neurological symptoms compared to the eastern population (Pardal et al., 2014; Torrez et al., 2015).

Much like *Centruroides*, members of the *Tityus* genus exhibit a diversity of NaTxS and KTxS, although we did not find venom characterization information for most species reported. Individual ion-channel toxins have been identified and/or functionally characterized from *T. caripitensis* (D'Suze et al., 2015), *T. discrepans* (Borges et al., 2006; Trejo et al., 2012; D'Suze et al., 2015), *T. pachyurus* (Barona et al., 2006; Guerrero-Vargas et al., 2012), *T. perijanensis* (D'Suze et al., 2015), *T. quirogae* (D'Suze et al., 2015), *T. trivittatus* (Coronas et al., 2003), and *T. zulianus* (Trejo et al., 2012; D'Suze et al., 2015).

Venom characterizations using transcriptomic, proteomic, or both approaches have also been completed for several *Tityus* species. NaTxS and KTxS were confirmed in the venom-gland transcriptome and proteome of *T. costatus* (Diego-García et al., 2005). In the venom-gland transcriptome and proteome of *T. stigmurus*, NaTxS, KTxS, AMPs and HYALS were present (Batista et al., 2007; Almeida et al., 2012). The venom of *T. metuendus* was proteomically characterized by Batista et al. (2018), who confirmed NaTxS, KTxS, MPs, HYALS, lectins and CRISPs. High-throughput transcriptomic methods were used to sequence the venom-gland transcriptome of *T. bahiensis*, which revealed a high abundance of MPs and confirmed the presence of NaTxS, KTxS, AMPs, Bpps, SPs, and CRISPs (de Oliveira et al., 2015). The most well-characterized *Tityus* venoms belong to *T. obscurus* and *T. serrulatus*. NaTxS, KTxS, AMPs, Bpps, HYALS, CRISPs, and MPs are among the venom components confirmed in the venom of *T. obscurus* using venom-gland transcriptomic and proteomic approaches (de Oliveira et al., 2018), some of which have been functionally characterized (Batista et al., 2000, 2002, 2004). The venom-gland transcriptome of *T. serrulatus* also revealed the presence of NaTxS, KTxS, AMPs and MPs (Alvarenga et al., 2012), most of which were later proteomically confirmed in the venom along with CRISPs and HYALS (de Oliveira et al., 2018). Several isolated *T. serrulatus* toxins have also been functionally characterized (Zoccal et al., 2011, 2013; Guo et al., 2013; Pucca et al., 2015b,c; 2016a, 2016b; Cerni et al., 2017).

### 3.2. Family Hemiscorpiidae

#### 3.2.1. Hemiscorpius

The *Hemiscorpius* genus currently consists of 15 species, several of which are found in Iran, Iraq, Oman, Saudi Arabia, Yemen, Pakistan, and the United Arab Emirates (Dehghani et al., 2018). We found two medically significant species belonging to the *Hemiscorpius* genus, *H. acanthocercus* and *H. lepturus*, although other members of this genus may be harmful to humans and mistakenly identified as *H. lepturus*. A sting from either *H. acanthocercus* or *H. lepturus* can be fatal, assigning them to sting class III, and they are each likely responsible for hundreds of envenomations annually (Table 3). Unlike medically significant *Buthidae* envenomations, the symptoms of *Hemiscorpius* envenomation often do not present immediately, and sometimes several days pass before severe symptoms begin, resulting in a delay in seeking available treatment that likely contributes to the severity of symptoms (Dehghani et al., 2018).

We could not find any confirmed venom toxins for *H. acanthocercus* in the literature, but the high-throughput venom-gland transcriptome of *H. lepturus* revealed a high abundance of AMPs, PLA2s, MPs, and HYALS, as well as a number of KTxS (Kazemi-Lomedasht et al., 2017). Several phospholipase D sequences were also identified in *H. lepturus*, one of which was isolated from the venom and was nearly identical in sequence and structure to the phospholipase D assumed to be responsible for high levels of toxicity and lethality of *Loxosceles* (brown recluse) spider venom (Torabi et al., 2017).

### 3.3. Family Scorpionidae

#### 3.3.1. Nebo

One species from the genus *Nebo*, *N. hierichonticus*, was identified as medically significant and is distributed throughout the Middle Eastern countries of Asia (Table 1). Reported life-threatening symptoms and fatalities caused by *N. hierichonticus* assigned this species to a sting class of III, although we only found a small number of cases reported (Table 3). We could not find any confirmed venom toxins for *N. hierichonticus* in the literature, however, we did find contradicting family classifications for the *Nebo* genus. Originally considered Scorpionidae, Prendini and Wheeler (2005) reclassified *Nebo* as belonging to the Diplocentridae family. Later that same year, Fet and Soleglad (2005) reclassified *Nebo* back to the Scorpionidae family. Recent literature has retained the Scorpionidae classification (Badry et al., 2018), so we have retained this position for the purpose of this review.

#### 3.4. Medical importance of harmless scorpions

Potential uses of scorpion venom toxins in medicine include AMPs in the development of novel antibiotics and effective malaria treatments (Conde et al., 2000; Zeng et al., 2005; Díaz et al., 2009; Fang et al., 2011; Guo et al., 2013; Harrison et al., 2014), Bpps in the treatment of hypotension (Camargo et al., 2012; Ortiz et al., 2015), and ion-channel toxins in the treatment of auto-immune diseases and cancer (Guo et al., 2013; Ortiz et al., 2015), including glioma tumors (DeBin and Strichartz, 1991; Deshane et al., 2003; Veisoh et al., 2007). An overwhelming majority of known scorpion species are considered harmless to humans and have not been considered as medically significant throughout the literature. Available venom characterizations of harmless scorpion species, however, suggest that these venoms may be just as medically useful as those that cause human harm, especially in the development of antibiotics and drugs with ion-channel specificity. In fact, many of the potential treatments listed above are based on toxins isolated from harmless species (Conde et al., 2000; Zeng et al., 2005; Fang et al., 2011; Harrison et al., 2014; Ortiz et al., 2015). Discussed here are a few more examples of well-characterized harmless scorpion venoms for the purpose of illustrating their potential in medicine.

Quintero-Hernández et al. (2015) characterized the venom-gland transcriptomes for four species belonging to the *Vaejovis* genus (Family Vaejovidae), *Vaejovis mexicanus*, *V. intrepidus*, *V. subcristatus*, and *V. punctatus*, which are largely distributed in Mexico. Although precursor coding sequences for NaTxS, KTxS, and CaTxS were detected in the *Vaejovis* transcriptomes, including potent KTxS shown to affect human potassium-channels (Gurrola et al., 2012), the most abundant transcripts present were those that code for AMPs of the non-disulfide bridge peptide (NDBP) class. The same group had previously isolated and functionally characterized some *Vaejovis* NDBPs and found them to be effective against both Gram-positive and Gram-negative bacterial strains (Ramírez-Carretero et al., 2012).

Santibáñez-López et al. (2017) characterized the venom-gland transcriptome and proteome of the Mexican scorpion, *Megacormus gertschi* (family Euscorpidae). Transcripts coding for NaTxS, KTxS, and CaTxS were present, however, these transcripts accounted for approximately 10% of the annotated toxin sequences combined. The NDBP class of AMPs also accounted for approximately 10% of annotated transcripts in *M. gertschi*, and the most abundant toxins present were enzymes such as MPs, SPs, PLA2s, and HYALS, where MPs alone accounted for nearly 20% of the annotated transcripts. Venom from *M. gertschi* was also demonstrated to be toxic in crickets and shrimp, and did not exhibit toxicity in mice, suggesting the toxins present are insect and crustacean specific.

Rokyta and Ward (2017) characterized the venom-gland transcriptome and proteome of *Hadrurus spadix*, a member of the Caraboctonidae family found in deserts of the eastern United States. The

most abundant and diverse toxins present in the venom of *H. spadix* were KTxS and AMPs (both NDBPs and cysteine-containing). A number of peptidases, PLA2s, and SPs were also detected, along with 66 venom protein (VP) transcripts with unknown function. The same group later characterized the venom-gland transcriptome and proteome of *Centruroides hentzi*, a harmless Buthidae species found in the southeastern United States (Ward et al., 2018). Similar to the harmful *Centruroides* species discussed above, the venom of *C. hentzi* primarily consisted of NaTxS and KTxS, although only one NaTx in *C. hentzi* was identified as mammal-specific.

### 3.5. Assessing the true medical significance of scorpions: problems and prospects

We identified 104 scorpion species considered medically significant in the literature, however, we could only assign 32 of these to a severe sting class of II or III based on specific symptoms attributed to these species. We found a handful of species that were implicated as being medically significant or harmful to humans in the literature, although documented case reports for these species indicated only class I symptoms. Of the 104 medically significant species identified, 68 were given an unknown sting class due to unverifiable symptoms or lack of symptoms reported. Several species were assigned an unknown sting classification because recent taxonomical updates have not yet been incorporated into the literature and it is unclear whether or not these reclassified species should be considered medically significant (*i.e.* *Buthus* and *Leiurus* species in particular).

The number of envenomations and severity of symptoms reported may depend on either human or scorpion population density (*i.e.* the likelihood of envenomation, Lourenço, 2018), as well as variation in sensitivity to envenomation, which may be age or health dependent (Pucca et al., 2015a; Hauke and Herzig, 2017). Several of the species presented here were assigned a sting classification based on symptoms reported in a single case study or in studies focused on envenomations in children and do not necessarily reflect a typical reaction in a healthy adult. It should therefore be noted that the term “medically significant” as it relates to scorpions in the literature does not necessarily equal a statistical significance in terms of the number of severe cases reported in proportion to the general population. Although thousands of deaths due to scorpion envenomation are reported annually, the majority of sting incidents, even by medically significant species, likely go unreported. Much larger and more comprehensive case studies that include both children and adults need to be completed to assess the true degree of harm a given species may have on humans. These studies should include proper specimen identification and those envenomated should be encouraged to bring in the offending scorpion (if safe to do so), especially in areas with high envenomation rates and diverse scorpion fauna. These same concerns have been addressed in the case of spider envenomations, leading to a strict standard of requirements for an apparent spider bite to be classified as such (Isbister and White, 2004). The importance of these standards was recently demonstrated by Stuber and Nentwig (2016), who indicated that only 22% of 134 medical spider bite case reports could be accurately verified. Although the reported species are not considered harmful to humans, the study of Australian scorpion stings by Isbister et al. (2003) is one such example of a more comprehensive case study that will ideally become the rule, rather than the exception, in the assessment of scorpion envenomations. Another consideration is the availability and access to scorpion envenomation treatment (Chippaux and Goyffon, 2008). Although treatment may effect the overall outcome of the patient (especially in the case of fatalities), any scorpion envenomation that requires medical treatment for survival or management of symptoms, even if only a single case study, should be classified as such (*i.e.* sting class II or III) to reflect the ability of the scorpion species to cause severe human harm and encourage those envenomated to seek appropriate treatment.

The reported medical significance of scorpions often implies that the

same toxins responsible for causing human harm may also be useful in therapeutic development. As such, we were surprised to find that only 54 of the 104 medically significant species we identified had undergone some form of venom characterization. Within these 54 species, most had partial characterizations with few individual toxins isolated and functionally characterized, and only 12 had more extensively characterized venoms using venom-gland transcriptomic and/or proteomic approaches. Although partial characterizations have contributed to therapeutic development (*i.e.* chlorotoxin, discussed above), these characterizations are likely missing key functional elements of the venom considering some toxins require activation by another to exhibit higher levels of toxicity (Undheim et al., 2014, 2015; Whittington et al., 2018). The complete characterization of venom, using venom-gland transcriptomic and proteomic approaches, will better ensure we are capturing the full potential of these powerful venoms. Additionally, many harmless species have already demonstrated their relevance in medicine, and they may hold the key in understanding the evolution of lethal toxins. Comparing harmful and harmless scorpion venoms will assist in our ability to decipher the differences in function, sequence, and abundance of similar toxins with varying consequences across multiple species. Therefore, more extensive venom characterizations should be a prioritized research effort in the study of both harmful and harmless scorpion species.

In the process of conducting our literature review, we found a seemingly large disconnect between those who encounter and/or provide treatment for scorpion envenomations (usually medical professionals), researchers who primarily focus on venom characterization or drug development, and researchers who focus on taxonomy and the identifying characteristics of scorpion species. Each of these contributing groups focus on a different area of scorpion research, all of which need to be integrated if we aim to grasp the true severity of scorpionism as well as develop medications from their venoms. Taxonomical classifications in particular are not readily or ubiquitously incorporated into other areas of the literature, making it difficult to decipher which species are truly medically significant or responsible for causing human harm. For the purposes of our review, we retained species names as reported in corresponding literature and noted where taxonomical updates should be applied, however, this does not account for species that were misidentified and therefore not accurately reported. Proper scorpion identification should be considered in any future scorpion studies encompassing all research areas and include the submission of voucher specimens for species confirmation. In cases where venom is not directly harvested by the researcher, as is common in studies focused on the characterization of isolated toxins, the researcher should clearly state the originating species and make every effort to obtain the venom from a source in which a voucher specimen has been submitted. The submission of voucher specimens would also leave room for future taxonomical updates that may impact results of previous studies. For example, *C. huichol* was classified as a new species after detailed examination of specimens thought to be *C. noxius* (Teruel et al., 2015). As a result, some envenomations that were attributed to *C. noxius* may actually belong to *C. huichol*.

## 4. Conclusions

In consideration of the many contributing factors discussed throughout this review, we have provided an updated, global list of scorpion species described as medically significant throughout the literature. Because scorpions are rapidly diversifying and new species are continuing to be described, the number of species, estimated sting frequencies, geographical boundaries, and sting classifications reported here should be considered a guideline only. Proper scorpion identification should become a priority in all future scorpion research, especially in the assessment of scorpionism and in work aimed at characterizing venoms for the purpose of drug development.

## Ethical statement

**Reporting standards:** The authors declare that our manuscript describes original research and every effort was made to ensure the accuracy of the results and the account.

**Data Access and Retention:** The authors declare that all data is reported and provided within the manuscript and there is no additional data to make available.

**Originality and Plagiarism:** The authors declare that our manuscript is an original work with proper citations as needed.

**Multiple, Redundant or Concurrent Publication:** The authors declare that the data and work described in our manuscript has not and will not be submitted for consideration to another journal.

**Acknowledgment of Sources:** The authors have provided proper acknowledgment of sources to the best of their abilities.

**Authorship of Paper:** All three authors of the manuscript made significant contributions, and no one making significant contributions was excluded from authorship. All three authors have seen and read the submitted version of the manuscript and have approved submission.

**Hazards and Human or Animal Subjects:** This review study did not involve the use of vertebrate animals.

**Disclosure and Conflicts of Interest:** The authors declare no conflicts of interest.

**Fundamental Errors in Published Works:** If a fundamental error or inaccuracy is discovered in the results described by our literature review, the authors will immediately notify the editor or publisher.

## Acknowledgments

We thank the editors and staff of *Toxicon*, with special thanks to Volker Herzig, for their invitation to contribute to the special edition on arthropod venoms. We also thank Darin Rokyta, Carl Whittington, and Michael Hogan for their comments on the manuscript, as well as to the reviewers for their thorough comments and suggestions. This material is based upon work supported by the National Science Foundation Graduate Research Fellowship Program under Grant No. 1449440. Any opinions, findings, and conclusions or recommendations expressed in this material are those of the authors and do not necessarily reflect the views of the National Science Foundation.

## Transparency document

Transparency document related to this article can be found online at <https://doi.org/10.1016/j.toxicon.2018.07.007>.

## References

- D'Suze, G., Castillo, C., Sevcik, C., Brazón, J., Malave, C., Hernandez, D., Zerpa, N., 2015. Scorpionism and dangerous species of Venezuela. In: *Scorpion Venoms*. Springer, pp. 273–298.
- Abdel-Mottaleb, Y., Clynen, E., Jalali, A., Bosmans, F., Vatanpour, H., Schoofs, L., Tytgat, J., 2006. The first potassium channel toxin from the venom of the Iranian scorpion *Odonthobuthus doriae*. *FEBS Lett.* 580, 6254–6258.
- Aboumaâd, B., Lahssaini, M., Tiger, A., Benhassain, S.M., 2014. Clinical comparison of scorpion envenomation by *Androctonus mauritanicus* and *Buthus occitanus* in children. *Toxicon* 90, 337–343.
- Aboutorabi, A., Naderi, N., Gholampour Pourbadiee, H., Zolfagharian, H., Vatanpour, H., 2016. Voltage-Gated sodium channels modulation by *Bothriatus schach* scorpion venom. *Iran. J. Pharm. Sci.* 12, 55–64.
- Adiguzel, S., 2010. In vivo and in vitro effects of scorpion venoms in Turkey: a mini-review. *J. Venom. Anim. Toxins Incl. Trop. Dis.* 16, 198–211.
- Akef, H., Kotb, N., Abo-Elmatty, D., Salem, S., 2017. Anti-proliferative effects of *Androctonus amoreuxi* scorpion and *Cerastes cerastes* snake venoms on human prostate cancer cells. *J. Cancer Prev.* 22, 40.
- Al-Sadoon, M., Jarrar, B., 2003. Epidemiological study of scorpion stings in Saudi Arabia between 1993 and 1997. *J. Venom. Anim. Toxins Incl. Trop. Dis.* 9, 54–64.
- Albuquerque, C. M. R. d., Porto, T.J., Amorim, M.L.P., Neto, S., de Lima, P., 2009. Scorpionism caused by *Tityus pusillus* Pocock, 1893 (scorpiones; Buthidae) in state of Pernambuco. *J. Braz. Soc. Trop. Med.* 42, 206–208.
- Almaaytah, A., Zhou, M., Wang, L., Chen, T., Walker, B., Shaw, C., 2012. Antimicrobial/cytolytic peptides from the venom of the North African scorpion, *Androctonus amoreuxi*: biochemical and functional characterization of natural peptides and a single site-substituted analog. *Peptides* 35, 291–299.
- Almeida, D.D., Scortecci, K.C., Kobashi, L.S., Agnez-Lima, L.F., Medeiros, S.R.B., Silva-Junior, A.A., Junqueira-de Azevedo, I.de.L.M., Fernandes-Pedrosa, M.de.F., 2012. Profiling the resting venom gland of the scorpion *Tityus stigmurus* through a transcriptomic survey. *BMC Genom.* 13, 362.
- Alvarenga, É.R., Mendes, T.M., Magalhães, B.F., Siqueira, F.F., Dantas, A.E., Barroca, T.M., Horta, C.C., Kalapothakis, E., 2012. Transcriptome analysis of the *Tityus serrulatus* scorpion venom gland. *Open J. Genet.* 2, 210–220.
- Amr, Z.S., Al Zoubi, R., Abdo, N., Hani, R.B., 2017. Scorpion stings in Jordan: an update. *Wilderness Environ. Med.* 28, 207–212.
- Annobil, S., 1993. Scorpion stings in children in the Asir province of Saudi Arabia. *J. Wilderness Med.* 4, 241–251.
- Badry, A., Younes, M., Sarhan, M.M., Saleh, M., 2018. On the scorpion fauna of Egypt, with an identification key (Arachnida: scorpiones). *Zool. Middle East* 64, 75–87.
- Bahloul, M., Ben Hamida, C., Chtourou, K., Ksibi, H., Dammak, H., Kallel, H., Chaari, A., Chelly, H., Guermazi, F., Rezik, N., Bouaziz, M., 2004. Evidence of myocardial ischaemia in severe scorpion envenomation. *Intensive Care Med.* 30, 461–467.
- Bahloul, M., Chaari, A., Khlaf-Bouaziz, N., Hergafi, L., Ksibi, H., Kallel, H., Chaari, A., Chelly, H., Hamida, C.B., Rezik, N., Bouaziz, M., 2005. Gastrointestinal manifestations in severe scorpion envenomation. *Gastroentérol. Clin. Biol.* 29, 1001–1005.
- Barona, J., Batista, C.V., Zamudio, F.Z., Gomez-Lagunas, F., Wanke, E., Otero, R., Possani, L.D., 2006. Proteomic analysis of the venom and characterization of toxins specific for Na<sup>+</sup>- and K<sup>+</sup>-channels from the Colombian scorpion *Tityus pachyurus*. *Biochim. Biophys. Acta BBA Proteins Proteomics* 1764, 76–84.
- Batista, C.V., Gómez-Lagunas, F., Lucas, S., Possani, L.D., 2000. Tc1, from *Tityus cambridgei*, is the first member of a new subfamily of scorpion toxin that blocks K<sup>+</sup>-channels. *FEBS Lett.* 486, 117–120.
- Batista, C.V., Zamudio, F.Z., Lucas, S., Fox, J.W., Frau, A., Prestipino, G., Possani, L.D., 2002. Scorpion toxins from *Tityus cambridgei* that affect Na<sup>+</sup>-channels. *Toxicon* 40, 557–562.
- Batista, C.V., del Pozo, L., Zamudio, F.Z., Contreras, S., Becerril, B., Wanke, E., Possani, L.D., 2004. Proteomics of the venom from the amazonian scorpion *Tityus cambridgei* and the role of prolines on mass spectrometry analysis of toxins. *J. Chromatogr. B* 803, 55–66.
- Batista, C., Roman-Gonzalez, S., Salas-Castillo, S., Zamudio, F., Gómez-Lagunas, F., Possani, L., 2007. Proteomic analysis of the venom from the scorpion *Tityus stigmurus*: biochemical and physiological comparison with other *Tityus* species. *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* 146, 147–157.
- Batista, C., Martins, J., Restano-Cassulini, R., Coronas, F., Zamudio, F., Procópio, R., Possani, L., 2018. Venom characterization of the amazonian scorpion *Tityus metuendus*. *Toxicon* 183, 51–58.
- Bavani, M.M., Rafinejad, J., Hanafi-Bojd, A.A., Oshaghi, M.A., Navidpour, S., Dabiri, F., Badakhshan, M., Ghorbani, E., Bagheri, M., 2017. Spatial distribution of medically important scorpions in North West of Iran. *J. Arthropod Borne Dis.* 11, 371.
- Bawaskar, H., Bawaskar, P., 1998. Indian red scorpion envenoming. *Indian J. Pediatr.* 65, 383–391.
- Bergman, N.J., 1997. Clinical description of *Parabuthus transvaalicus* scorpionism in Zimbabwe. *Toxicon* 35, 759–771.
- Bertani, R., Bonini, R.K., Toda, M.M., Isa, L.S., Vono, J., Alvarez Figueiredo, M. R. d. S., Ferraz, S.C., 2018. Alien scorpions in the municipality of São Paulo, Brazil—evidence of successful establishment of *Tityus stigmurus* (Thorell, 1876) and first records of *Broteochactas parvulus* Pocock, 1897, and *Jaguajir rochae* (Borelli, 1910). *BiolInvasions Rec.* 7, 89–94.
- Borges, A., García, C.C., Lugo, E., Alfonso, M.J., Jowers, M.J., den Camp, H.J.O., 2006. Diversity of long-chain toxins in *Tityus zulianus* and *Tityus discrepans* venoms (Scorpiones, Buthidae): molecular, immunological, and mass spectral analyses. *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* 142, 240–252.
- Borges, A., Rojas-Runjaic, F.J., Diez, N., Faks, J.G., den Camp, H.J.O., De Sousa, L., 2010. Envenomation by the scorpion *Tityus breweri* in the Guayana Shield, Venezuela: report of a case, efficacy and reactivity of antivenom, and proposal for a toxinological partitioning of the Venezuelan scorpion fauna. *Wilderness Environ. Med.* 21, 282–290.
- Borges, A., den Camp, H.J.O., De Sanctis, J.B., 2011. Specific activation of human neutrophils by scorpion venom: a flow cytometry assessment. *Toxicol. Vitro* 25, 358–367.
- Borges, A., Miranda, R., Pascale, J., 2012. Scorpionism in Central America, with special reference to the case of Panama. *J. Venom. Anim. Toxins Incl. Trop. Dis.* 18, 130–143.
- Borges, A., Morales, M., Loo, W., Delgado, M., 2015. Scorpionism in Ecuador: first report of severe and fatal envenoming cases from northern Manabí by *Tityus asthenes* Pocock. *Toxicon* 105, 56–61.
- Brasil, 2009. Manual de controle de escorpiões, Ministério da Saúde Brasília.
- Bucarechi, F., Fernandes, L.C., Fernandes, C.B., Branco, M.M., Prado, C.C., Vieira, R.J., De Capitani, E.M., Hyslop, S., 2014. Clinical consequences of *Tityus bahiensis* and *Tityus serrulatus* scorpion stings in the region of Campinas, southeastern Brazil. *Toxicon* 89, 17–25.
- Bush, S.P., 1999. Envenomation by the scorpion (*Centruroides limbatus*) outside its natural range and recognition of medically important scorpions. *Wilderness Environ. Med.* 10, 161–164.
- Caliskan, F., Quintero-Hernández, V., Restano-Cassulini, R., Batista, C., Zamudio, F., Coronas, F., Possani, L., 2012. Turkish scorpion *Buthacus macrocentrus*: general characterization of the venom and description of Bu1, a potent mammalian Na<sup>+</sup>-channel  $\alpha$ -toxin. *Toxicon* 59, 408–415.
- Caliskan, F., Quintero-Hernández, V., Restano-Cassulini, R., Coronas-Valderrama, F.I., Corzo, G., Possani, L.D., 2013. Molecular cloning and biochemical characterization of the first Na<sup>+</sup>-channel  $\alpha$ -type toxin peptide (Acra4) from *Androctonus crassicauda* scorpion venom. *Biochimie* 95, 1216–1222.

- Camargo, A.C., Ianzer, D., Guerreiro, J.R., Serrano, S.M., 2012. Bradykinin-potentiating peptides: beyond captopril. *Toxicon* 59, 516–523.
- Carmo, A., Oliveira-Mendes, B., Horta, C., Magalhães, B., Dantas, A., Chaves, L., Chavez-Olortegui, C., Kalapothakis, E., 2014. Molecular and functional characterization of metalloserulases, new metalloproteases from the *Tityus serrulatus* venom gland. *Toxicon* 90, 45–55.
- Castle, N.A., Strong, P.N., 1986. Identification of two toxins from scorpion (*Leiurus quinquestriatus*) venom which block distinct classes of calcium-activated potassium channel. *FEBS Lett.* 209, 117–121.
- Cerni, F.A., Pucca, M.B., Zoccal, K.F., Frantz, F.G., Faccioli, L.H., Arantes, E.C., 2017. Expanding biological activities of Ts19 Frag-II toxin: insights into IL-17 production. *Toxicon* 134, 18–25.
- Cesaretti, Y., Ozkan, O., 2010. Scorpion stings in Turkey: epidemiological and clinical aspects between the years 1995 and 2004. *Rev. do Inst. Med. Trop. São Paulo* 52, 215–220.
- Chakroun-Walha, O., Karray, R., Jerbi, M., Nasri, A., Issaoui, F., Amine, B.R., Bahloul, M., Bouazziz, M., Ksibi, H., Rekiq, N., 2018. Update on the epidemiology of scorpion envenomation in the south of Tunisia. *Wilderness Environ. Med.* 29, 29–35.
- Chen, T., Polan, R., Kwok, H., O'Kane, E.J., Bjourson, A.J., Shaw, C., 2003. Isolation of scorpion (*Androctonus amoreuxi*) putative alpha neurotoxins and parallel cloning of their respective cDNAs from a single sample of venom. *Regul. Pept.* 115, 115–121.
- Chen, T., Walker, B., Zhou, M., Shaw, C., 2005. Molecular cloning of a novel putative potassium channel-blocking neurotoxin from the venom of the North African scorpion, *Androctonus amoreuxi*. *Peptides* 26, 731–736.
- Chippaux, J.-P., Goyffon, M., 2008. Epidemiology of scorpionism: a global appraisal. *Acta Trop.* 107, 71–79.
- Chowell, G., Díaz-Dueñas, P., Bustos-Saldaña, R., Mireles, A.A., Fet, V., 2006. Epidemiological and clinical characteristics of scorpionism in Colima, Mexico (2000–2001). *Toxicon* 47, 753–758.
- Coelho, P., Sousa, P., Harris, D., van der Meijden, A., 2014. Deep intraspecific divergences in the medically relevant fat-tailed scorpions (*Androctonus*, scorpiones). *Acta Trop.* 134, 43–51.
- Coelho, J.S., Ishikawa, E.A.Y., Dos Santos, P.R.S.G., de Oliveira Pardal, P.P., 2016. Scorpionism by *Tityus silvestris* in eastern Brazilian amazon. *J. Venom. Anim. Toxins Incl. Trop. Dis.* 22, 24.
- Conde, R., Zamudio, F.Z., Rodríguez, M.H., Possani, L.D., 2000. Scorpine, an anti-malaria and anti-bacterial agent purified from scorpion venom. *FEBS Lett.* 471, 165–168.
- Corona, M., Valdez-Cruz, N., Merino, E., Zurita, M., Possani, L., 2001. Genes and peptides from the scorpion *Centruroides sculpturatus* (Ewing), that recognize Na<sup>+</sup>-channels. *Toxicon* 39, 1893–1898.
- Coronas, F.V., de Roodt, A.R., Olamendi-Portugal, T., Zamudio, F.Z., Batista, C.V., Gómez-Lagunas, F., Possani, L.D., 2003. Disulfide bridges and blockage of Shaker B K<sup>+</sup>-channels by another butantoxin peptide purified from the Argentinean scorpion *Tityus trivittatus*. *Toxicon* 41, 173–179.
- Corzo, G., Papp, F., Varga, Z., Barraza, O., Espino-Solis, P.G., de la Vega, R.C.R., Gaspar, R., Panyi, G., Possani, L.D., 2008. A selective blocker of Kv1. 2 and Kv1. 3 potassium channels from the venom of the scorpion *Centruroides suffusus suffusus*. *Biochem. Pharmacol.* 76, 1142–1154.
- Costa, C. L. S. d. O., Sampaio, N. F. Fé, I., Tadei, W.P., 2016. A profile of scorpionism, including the species of scorpions involved, in the State of Amazonas, Brazil. *Rev. Soc. Bras. Med. Trop.* 49, 376–379.
- Cupo, P., Hering, S.E., 2002. Cardiac troponin I release after severe scorpion envenoming by *Tityus serrulatus*. *Toxicon* 40, 823–830.
- Daisley, H., Alexander, D., Pitt-Miller, P., 1999. Acute myocarditis following *Tityus trinitatis* envenoming: morphological and pathophysiological characteristics. *Toxicon* 37, 159–165.
- Dardevet, L., Rani, D., Aziz, T.A.E., Bazin, I., Sabatier, J.-M., Fadl, M., Brambilla, E., De Waard, M., 2015. Chlorotoxin: a helpful natural scorpion peptide to diagnose glioma and fight tumor invasion. *Toxins* 7, 1079–1101.
- DeBin, J., Strichartz, G., 1991. Chloride channel inhibition by the venom of the scorpion *Leiurus quinquestriatus*. *Toxicon* 29, 1403–1408.
- Debont, T., Swerts, A., van der Walt, J.J., Müller, G.J., Verdonck, F., Daenens, P., Tytgat, J., 1998. Comparison and characterization of the venoms of three *Parabuthus* scorpion species occurring in southern Africa. *Toxicon* 36, 341–352.
- Dehesa-Dávila, M., Ramfrez, A.N., Zamudio, F.Z., Gurrola-Briones, G., Liévano, A., Darzon, A., Possani, L.D., 1996. Structural and functional comparison of toxins from the venom of the scorpions *Centruroides infamatus infamatus*, *Centruroides limpidus limpidus* and *Centruroides noxius*. *Comp. Biochem. Physiol. Part B Biochem. Mol. Biol.* 113, 331–339.
- Dehghani, R., Fathi, B., 2012. Scorpion sting in Iran: a review. *Toxicon* 60, 919–933.
- Dehghani, R., Djadid, N.D., Shahbazzadeh, D., Bigdelli, S., 2009. Introducing *Compsothous matthiesseni* (Birula, 1905) scorpion as one of the major stinging scorpions in Khuzestan, Iran. *Toxicon* 54, 272–275.
- Dehghani, R., Kamiabi, F., Mohammadi, M., 2018. Scorpionism by *Hemiscorpius spp.* in Iran: a review. *J. Venom. Anim. Toxins Incl. Trop. Dis.* 24, 8.
- Deshane, J., Garner, C.C., Sontheimer, H., 2003. Chlorotoxin inhibits glioma cell invasion via matrix metalloproteinase-2. *J. Biol. Chem.* 278, 4135–4144.
- Deshpande, S., Alex, A., Jagannadham, M., Rao, G., Tiwari, A., 2005. Identification of a novel pulmonary oedema producing toxin from Indian red scorpion (*Mesobuthus tamulus*) venom. *Toxicon* 45, 735–743.
- Dias, S.C., Candido, D.M., Brescovit, A.D., 2006. Scorpions from mata do Buraquinho, João Pessoa, Paraíba, Brazil, with ecological notes on a population of *Ananteris mawryi* Lourenço (scorpiones, Buthidae). *Rev. Bras. Zool.* 23, 707–710.
- Díaz, P., D'suze, G., Salazar, V., Sevcik, C., Shannon, J.D., Sherman, N.E., Fox, J.W., 2009. Antibacterial activity of six novel peptides from *Tityus discrepans* scorpion venom. A fluorescent probe study of microbial membrane Na<sup>+</sup> permeability changes. *Toxicon* 54, 802–817.
- Diego-García, E., Batista, C.V., García-Gómez, B.I., Lucas, S., Candido, D.M., Gómez-Lagunas, F., Possani, L.D., 2005. The Brazilian scorpion *Tityus costatus* karsch: genes, peptides and function. *Toxicon* 45, 273–283.
- Diego-García, E., Peigneur, S., Clynen, E., Marien, T., Czech, L., Schoofs, L., Tytgat, J., 2012. Molecular diversity of the telson and venom components from *Pandinus cavimanus* (Scorpionidae Latreille 1802): transcriptome, venomomics and function. *Proteomics* 12, 313–328.
- Diego-García, E., Peigneur, S., Debaveye, S., Gheldof, E., Tytgat, J., Caliskan, F., 2013. Novel potassium channel blocker venom peptides from *Mesobuthus gibbosus* (Scorpiones: Buthidae). *Toxicon* 61, 72–82.
- Diego-García, E., Caliskan, F., Tytgat, J., 2014. The mediterranean scorpion *Mesobuthus gibbosus* (Scorpiones, Buthidae): transcriptome analysis and organization of the genome encoding chlorotoxin-like peptides. *BMC Genom.* 15, 295.
- Dokur, M., Dogan, M., Yagmur, E.A., 2017. Scorpion-related cardiomyopathy and acute pulmonary edema in a child who is stung by *Leiurus abduallahbayrami*. *Turk. J. Emerg. Med.* 17, 104–108.
- Du, Q., Hou, X., Wang, L., Zhang, Y., Xi, X., Wang, H., Zhou, M., Duan, J., Wei, M., Chen, T., et al., 2015. AaeAP1 and AaeAP2: novel antimicrobial peptides from the venom of the scorpion, *Androctonus aeneas*: structural characterisation, molecular cloning of biosynthetic precursor-encoding cDNAs and engineering of analogues with enhanced antimicrobial and anticancer activities. *Toxins* 7, 219–237.
- Dudina, E.E., Korolkova, Y.V., Bocharova, N.E., Koshelev, S.G., Egorov, T.A., Huys, I., Tytgat, J., Grishin, E.V., 2001. Osk2, a new selective inhibitor of Kv1. 2 potassium channels purified from the venom of the scorpion *Orthochirus scrobiculosus*. *Biochem. Biophys. Res. Commun.* 286, 841–847.
- Dyason, K., Brandt, W., Prendini, L., Verdonck, F., Tytgat, J., Plessis, J. d., Müller, G., Walt, J. v. d., 2002. Determination of species-specific components in the venom of *Parabuthus* scorpions from southern Africa using matrix-assisted laser desorption time-of-flight mass spectrometry. *Rapid Commun. Mass Spectrom.* 16, 768–773.
- D'suze, G., Moncada, S., González, C., Sevcik, C., Aguilar, V., Alagón, A., 2003. Relationship between plasmatic levels of various cytokines, tumour necrosis factor, enzymes, glucose and venom concentration following *Tityus* scorpion sting. *Toxicon* 41, 367–375.
- Emerich, B.L., De Lima, M.E., Martin-Eauclaire, M.-F., Bougis, P.E., 2017. Comparative analyses and implications for antivenom serotherapy of four Moroccan scorpion *Buthus occitanus* venoms: subspecies *tunetanus*, *paris*, *malhommei*, and *mardochei*. *Toxicon* 149, 26–36.
- Erickson, T.B., Cheema, N., 2017. Arthropod envenomation in North America. *Emerg. Med. Clin.* 35, 355–375.
- Esposito, L.A., Yamaguti, H.Y., Souza, C.A., Pinto-Da-Rocha, R., Prendini, L., 2017. Systematic revision of the neotropical club-tailed scorpions, physoctonus, rhopalurus, and troglorhopalurus, revalidation of heteroctenus, and descriptions of two new genera and three new species (Buthidae: rhopalurusinae). *Bull. Am. Mus. Nat. Hist.* 1–136.
- Estrada, G., Garcia, B.I., Schiavon, E., Ortiz, E., Cestele, S., Wanke, E., Possani, L.D., Corzo, G., 2007. Four disulfide-bridged scorpion beta neurotoxin CssiI: heterologous expression and proper folding in vitro. *Biochim. Biophys. Acta BBA Gen. Subj.* 1770, 1161–1168.
- Estrada-Gómez, S., Gomez-Rave, L., Vargas-Muñoz, L.J., van der Meijden, A., 2017. Characterizing the biological and biochemical profile of six different scorpion venoms from the Buthidae and Scorpionidae family. *Toxicon* 130, 104–115.
- Fang, W., Vega-Rodríguez, J., Ghosh, A.K., Jacobs-Lorena, M., Kang, A., Leger, R.J.S., 2011. Development of transgenic fungi that kill human malaria parasites in mosquitoes. *Science* 331, 1074–1077.
- Fet, V., Soleglad, M.E., 2005. Contributions to scorpion systematics. I. On recent changes in high-level taxonomy. *Euscorpius* 2005, 1–13.
- Fet, V., Sissom, W.D., Lowe, G., Braunwalder, M.E., et al., 2000. Catalog of the Scorpions of the World (1758–1998). New York Entomological Society.
- Fet, V., Kovařík, F., Gantenbein, B., Kaiser, R.C., Stewart, A.K., Graham, M.R., 2018. Revision of the *Mesobuthus caucasicus* complex from Central Asia, with descriptions of six new species (Scorpiones: Buthidae). *Euscorpius* 2018, 1–77.
- Gao, B., Peigneur, S., Tytgat, J., Zhu, S., 2010a. A potent potassium channel blocker from *Mesobuthus eupeus* scorpion venom. *Biochimie* 92, 1847–1853.
- Gao, B., Xu, J., del Carmen Rodriguez, M., Lanz-Mendoza, H., Hernández-Rivas, R., Du, W., Zhu, S., 2010b. Characterization of two linear cationic antimalarial peptides in the scorpion *Mesobuthus eupeus*. *Biochimie* 92, 350–359.
- García, M.L., García-Calvo, M., Hidalgo, P., Lee, A., MacKinnon, R., 1994. Purification and characterization of three inhibitors of voltage-dependent K<sup>+</sup>-channels from *Leiurus quinquestriatus* var. *hebraeus* venom. *Biochemistry* 33, 6834–6839.
- García-Calvo, M., Leonard, R., Novick, J., Stevens, S., Schmalhofer, W., Kaczorowski, G., García, M., 1993. Purification, characterization, and biosynthesis of margatoxin, a component of *Centruroides margaritatus* venom that selectively inhibits voltage-dependent potassium channels. *J. Biol. Chem.* 268, 18866–18874.
- Ghalim, N., El-Hafny, B., Sebt, F., Heikel, J., Lazar, N., Moustani, R., Benslimane, A., 2000. Scorpion envenomation and serotherapy in Morocco. *Am. J. Trop. Med. Hyg.* 62, 277–283.
- Goyffon, M., Dabo, A., Coulibaly, S., Togo, G., Chippaux, J.-P., 2012. Dangerous scorpion fauna of Mali. *J. Venom. Anim. Toxins Incl. Trop. Dis.* 18, 361–368.
- Guerrero-Vargas, J.A., Mourão, C.B., Quintero-Hernández, V., Possani, L.D., Schwartz, E.F., 2012. Identification and phylogenetic analysis of *Tityus pachyurus* and *Tityus obscurus* novel putative Na<sup>+</sup>-channel scorpion toxins. *PLoS One* 7, e30478.
- Gummin, D.D., Mowry, J.B., Spyker, D.A., Brooks, D.E., Fraser, M.O., Banner, W., 2017. 2016 annual report of the American association of poison control centers' national poison data System (NPDS): 34th annual report. *Clin. Toxicol.* 55, 1072–1254.
- Guo, X., Ma, C., Du, Q., Wei, R., Wang, L., Zhou, M., Chen, T., Shaw, C., 2013. Two

- peptides, TsAP-1 and TsAP-2, from the venom of the Brazilian yellow scorpion, *Tityus serrulatus*: evaluation of their antimicrobial and anticancer activities. *Biochimie* 95, 1784–1794.
- Gurrola, G.B., Hernández-López, R.A., Rodríguez de la Vega, R.C., Varga, Z., Batista, C.V., Salas-Castillo, S.P., Panyi, G., del Río-Portilla, F., Possani, L.D., 2012. Structure, function, and chemical synthesis of *Vaejovis mexicanus* peptide 24: a novel potent blocker of Kv1.3 potassium channels of human T lymphocytes. *Biochemistry* 51, 4049–4061.
- Harrison, P.L., Abdel-Rahman, M.A., Miller, K., Strong, P.N., 2014. Antimicrobial peptides from scorpion venoms. *Toxicon* 88, 115–137.
- Hauke, T.J., Herzog, V., 2017. Dangerous arachnids—Fake news or reality? *Toxicon* 138, 173–183.
- He, Y., Zhao, R., Di, Z., Li, Z., Xu, X., Hong, W., Wu, Y., Zhao, H., Li, W., Cao, Z., 2013. Molecular diversity of the Chaerilidae venom peptides reveals the dynamic evolution of scorpion venom components from Buthidae to non-Buthidae. *J. Proteom.* 89, 1–14.
- El Hidan, M.A., Touloun, O., Boumezzough, A., 2017. Spatial relationship between environmental factors and scorpion distribution in Morocco. *J. Entomol. Zool. Stud.* 5, 674–678.
- El Hidan, M.A., Touloun, O., Bouazza, A., Laaradia, M.A., Boumezzough, A., 2018. *Androctonus* genus species in arid regions: ecological niche models, geographical distributions, and envenomation risk. *Vet. World* 11, 286.
- Himou, R., Soulaymani, A., Mokhtari, A., Arfaoui, A., Eloufir, G., Semlali, I., Soulaymani Bencheikh, R., 2008. Risk factors caused by scorpion stings and envenomations in the province of Kelâa Des Sraghna (Morocco). *J. Venom. Anim. Toxins Incl. Trop. Dis.* 14, 628–640.
- Isbister, G.K., White, J., 2004. Clinical consequences of spider bites: recent advances in our understanding. *Toxicon* 43, 477–492.
- Isbister, G.K., Volschenk, E.S., Balit, C.R., Harvey, M.S., 2003. Australian scorpion stings: a prospective study of definite stings. *Toxicon* 41, 877–883.
- Izquierdo, L.M., Buitrago, J.R.R., 2012. Cardiovascular dysfunction and pulmonary edema secondary to severe envenoming by *Tityus pachyurus* sting. Case report. *Toxicon* 60, 603–606.
- Jalali, A., Bosmans, F., Amininasab, M., Clynen, E., Cuyper, E., Zaremirakabadi, A., Sarbolouki, M.-N., Schoofs, L., Vatanpour, H., Tytgat, J., 2005. OD1, the first toxin isolated from the venom of the scorpion *Odonthobuthus doriae* active on voltage-gated Na<sup>+</sup> channels. *FEBS Lett.* 579, 4181–4186.
- Jalali, A., Pipelzadeh, M.H., Sayedian, R., Rowan, E., 2010. A review of epidemiological, clinical and in vitro physiological studies of envenomation by the scorpion *Hemiscorpius lepturus* (Hemiscorpiidae) in Iran. *Toxicon* 55, 173–179.
- Jiménez-Jiménez, M., Palacios-Cardiel, C., 2010. Scorpions of desert oases in the southern Baja California Peninsula. *J. Arid Environ.* 74, 70–74.
- Kaltsas, D., Stathi, I., Mylonas, M., 2008. The foraging activity of *Mesobuthus gibbosus* (Scorpiones: Buthidae) in central and south Aegean archipelago. *J. Nat. Hist.* 42, 513–527.
- Kang, A.M., Brooks, D.E., 2017. Nationwide scorpion exposures reported to US poison control centers from 2005 to 2015. *J. Med. Toxicol.* 13, 158–165.
- Karataş, A., Karataş, A., 2003. *Mesobuthus eupeus* (CL Koch, 1839) (scorpiones: Buthidae) in Turkey. *Euscorpius* 2003, 1–6.
- Kazemi-Lomedasht, F., Khalaj, V., Bagheri, K.P., Behdani, M., Shahbazzadeh, D., 2017. The first report on transcriptome analysis of the venom gland of Iranian scorpion, *Hemiscorpius lepturus*. *Toxicon* 125, 123–130.
- Khalil, N., Yağmur, E.A., 2010. *Leiurus abdullahbayrami* (Scorpiones: Buthidae), a new species for the scorpion fauna of Syria. *Serket* 12, 1–6.
- Khattabi, A., Soulaymani-Bencheikh, R., Achour, S., Salmi, L.-R., 2011. Classification of clinical consequences of scorpion stings: consensus development. *Trans. R. Soc. Trop. Med. Hyg.* 105, 364–369.
- Koschak, A., Bugianesi, R.M., Mitterdorfer, J., Kaczowski, G.J., Garcia, M.L., Knaus, H.-G., 1998. Subunit composition of Brain voltage-gated potassium channels determined by Hongotoxin-1, a novel peptide derived from *Centruroides limbatus* venom. *J. Biol. Chem.* 273, 2639–2644.
- Kovařík, F., 2007. A revision of the genus *Hottentotta* Birula, 1908, with descriptions of four new species (Scorpiones, Buthidae). *Euscorpius* 2007, 1–107.
- Kularatne, S.A., Dinamithra, N.P., Sivansuthan, S., Weerakoon, K.G., Thillaimpalam, B., Kalyanasundaram, V., Ranawana, K.B., 2015. Clinic-epidemiology of stings and envenoming of *Hottentotta tamulus* (scorpiones: Buthidae), the indian red scorpion from Jaffna Peninsula in northern Sri Lanka. *Toxicon* 93, 85–89.
- Kumar, R.B., Priya, B.S., Suresh, M.X., 2015. In silico analysis of potential inhibitors of Ca<sup>2+</sup> activated K<sup>+</sup> channel blocker, Charybdotoxin-C from *Leiurus quinquestriatus hebraeus* through molecular docking and dynamics studies. *Indian J. Pharmacol.* 47, 280.
- Kuzmenkov, A.I., Vassilevski, A.A., Kudryashova, K.S., Nekrasova, O.V., Peigneur, S., Tytgat, J., Feofanov, A.V., Kirpichnikov, M.P., Grishin, E.V., 2015. Variability of potassium channel blockers in *Mesobuthus eupeus* scorpion venom with focus on Kv1.1 an integrated transcriptomic and proteomic study. *J. Biol. Chem.* 290, 12195–12209.
- Laaradia, M.A., El Hidan, M.A., Marhoume, F., Boumeja, B., Oufquir, S., Sokar, Z., Boumezzough, A., Chait, A., 2018. *Buthus lienhardi* venom and pathophysiological effects at the histological, hematological, biochemical and motor skills levels. *Toxicon* 146, 106–113.
- Laraba-Djebari, F., Legros, C., Crest, M., Ceard, B., Romi, R., Mansuelle, P., Jacquet, G., Van Rietschoten, J., Gola, M., Rochat, H., 1994. The kaliotoxin family enlarged. purification, characterization, and precursor nucleotide sequence of KTx2 from *Androctonus australis* venom. *J. Biol. Chem.* 269, 32835–32843.
- Lebreton, F., Delepiere, M., Ramirez, A., Balderas, C., Possani, L., 1994. Primary and NMR three-dimensional structure determination of a novel crustacean toxin from the venom of the scorpion *Centruroides limpidus limpidus* Karsch. *Biochemistry* 33, 11135–11149.
- Lira-da Silva, R.M., Amorim, A. M. d., Brazil, T.K., 2000. Envenomation by *Tityus stigmurus* (scorpiones; Buthidae) in Bahia, Brazil. *Rev. Soc. Bras. Med. Trop.* 33, 239–245.
- Lobo, R.A., Galdoni, P.A.M., Souza, C. A. R. d., Medeiros, C. R. d., 2011. Accident caused by *Centruroides testaceus* (DeGeer, 1778) (scorpiones, Buthidae), native to the caribbean, in Brazilian airport. *Rev. Soc. Bras. Med. Trop.* 44, 789–791.
- Lourenço, W.R., Pezier, A., 2002. Taxonomic consideration of the genus *Odonthobuthus* Vachon (Scorpiones, Buthidae), with description of a new species. *Rev. Suisse Zool.* 109, 115–125.
- Lourenço, W.R., 2008. Parthenogenesis in scorpions: some history-new data. *J. Venom. Anim. Toxins Incl. Trop. Dis.* 14, 19–44.
- Lourenço, W.R., 2015. What do we know about some of the most conspicuous scorpion species of the genus *Tityus*? a historical approach. *J. Venom. Anim. Toxins Incl. Trop. Dis.* 21, 20.
- Lourenço, W.R., 2016. Scorpion incidents, misidentification cases and possible implications for the final interpretation of results. *J. Venom. Anim. Toxins Incl. Trop. Dis.* 22, 21.
- Lourenço, W., 2017. Scorpions from Brazilian Amazonia, with a description of two new species from Serra da Mocidade National Park in the State of Roraima (Scorpiones: Buthidae, Chactidae). *Arachnida Riv. Aracnol. Ital.* 12, 2–17.
- Lourenço, W.R., 2018. The evolution and distribution of noxious species of scorpions (Arachnida: Scorpiones). *J. Venom. Anim. Toxins Incl. Trop. Dis.* 24, 1.
- Lourenço, W.R., Leguin, E.-A., 2008. The true identity of *Scorpio (Atreus) obscurus* Gervais, 1843 (scorpiones, Buthidae). *Euscorpius* 2008, 1–9.
- Lourenço, W., Rossi, A., 2016. One more African species of the genus *Leiurus* Ehrenberg, 1828 (scorpiones: Buthidae) from Somalia. *Arachnida* 6, 21–31.
- Lourenço, W., da Silva, E.A., 2007. New evidence for a disrupted distribution pattern of the *Tityus confluens* complex, with the description of a new species from the state of Pará, Brazil (Scorpiones, Buthidae). *Amazoniana* 19, 77–86.
- Lourenço, W.R., Qi, J.-X., Cloudsley-Thompson, J.L., 2006. The African species of the genus *Leiurus* Ehrenberg, 1828. *Bol. Soc. Entomol. Aragon.* 1, 97–101.
- Lourenço, W., Kourim, M., Sadine, S., 2018. Scorpions from the region of tamanrasset, Algeria. Part II. A new African species of the genus *Leiurus* Ehrenberg, 1828 (scorpiones: Buthidae). *Arachnida Riv. Aracnol. Ital.* 16, 3–14.
- Lowe, G., Yağmur, E.A., Kovařík, F., 2014. A review of the genus *Leiurus* Ehrenberg, 1828 (Scorpiones: Buthidae) with description of four new species from the Arabian Peninsula. *Euscorpius* 2014, 1–129.
- Luna-Ramírez, K., Quintero-Hernández, V., Juárez-González, V.R., Possani, L.D., 2015. Whole transcriptome of the venom gland from *Urodacus yaschenko* scorpion. *PLoS One* 10, e0127883.
- Ma, Y., Zhao, R., He, Y., Li, S., Liu, J., Wu, Y., Cao, Z., Li, W., 2009. Transcriptome analysis of the venom gland of the scorpion *Scorpiops jendeki*: implication for the evolution of the scorpion venom arsenal. *BMC Genom.* 10, 290.
- Marinkelle, C., Stahnke, H., 1965. Toxicological and clinical studies on *Centruroides margaritatus* (Gervais), a common scorpion in western Colombia. *J. Med. Entomol.* 2, 197–199.
- Martin, M., Perez, L. G. y, El Ayeb, M., Kopeyan, C., Bechis, G., Jover, E., Rochat, H., 1987. Purification and chemical and biological characterizations of seven toxins from the Mexican scorpion, *Centruroides suffusus suffusus*. *J. Biol. Chem.* 262, 4452–4459.
- Martin-Eauclaire, M.-F., Bougis, P.E., 2012. Potassium channels blockers from the venom of *Androctonus mauretanicus mauretanicus*. *J. Toxicol.* <https://doi.org/10.1155/2012/103608>.
- Martin-Eauclaire, M.-F., Bosmans, F., Céard, B., Diocot, S., Bougis, P.E., 2014. A first exploration of the venom of the *Buthus occitanus* scorpion found in southern France. *Toxicon* 79, 55–63.
- van der Meijden, A., Koch, B., van der Valk, T., Vargas-Muñoz, L.J., Estrada-Gómez, S., 2017. Target-specificity in scorpions; comparing lethality of scorpion venoms across arthropods and vertebrates. *Toxins* 9, 312.
- Meki, A.-R.M., Nassar, A.Y., Rochat, H., 1995. A bradykinin-potentiating peptide (peptide K12) isolated from the venom of Egyptian scorpion *Buthus occitanus*. *Peptides* 16, 1359–1365.
- Monteiro, W.M., de Oliveira, S.S., Pivoto, G., Alves, E.C., Sachett, J.d.A.G., Alexandre, C.N., Fé, N.F., Guerra, M.d.G.V.B., da Silva, I.M., Tavares, A.M., et al., 2016. Scorpion envenoming caused by *Tityus cf. silvestris* evolving with severe muscle spasms in the Brazilian Amazon. *Toxicon* 119, 266–269.
- Mouhat, S., Visan, V., Ananthkrishnan, S., Wulff, H., Andreotti, N., Grissmer, S., Darbon, H., De Waard, M., Sabatier, J.-M., 2005. K<sup>+</sup> channel types targeted by synthetic OSK1, a toxin from *Orthochirus scrobiculosus* scorpion venom. *Biochem. J.* 385, 95–104.
- Mowry, J.B., Spyker, D.A., Brooks, D.E., McMillan, N., Schauben, J.L., 2015. 2014 annual report of the American association of poison control centers' national poison data System (NPPDS): 32nd annual report. *Clin. Toxicol.* 53, 962–1147.
- Mowry, J.B., Spyker, D.A., Brooks, D.E., Zimmerman, A., Schauben, J.L., 2016. 2015 annual report of the American association of poison control centers' national poison data System (NPPDS): 33rd annual report. *Clin. Toxicol.* 54, 924–1109.
- Müller, G., 1993. Scorpionism in South Africa: a report of 42 serious scorpion envenomations. *S. Afr. Med. J.* 83, 405–411.
- Naderi-Soorki, M., Galehdari, H., Baradaran, M., Jalali, A., 2016. First venom gland transcriptomic analysis of Iranian yellow scorpion *Odonthobuthus doriae* with some new findings. *Toxicon* 120, 69–77.
- Navidpour, S., 2015. An annotated checklist of scorpions in south and southwestern parts of Iran. *Int. J. Fauna Biol. Stud.* 2, 9–15.
- Nejati, J., Mozafari, E., Saghafipour, A., Kiyani, M., 2014. Scorpion fauna and epidemiological aspects of scorpionism in southeastern Iran. *Asian Pac. J. Trop. Biomed.* 4, S217–S221.

- Ojanguren-Affilastro, A.A., Adilardi, R.S., Mattoni, C.I., Ramírez, M.J., Ceccarelli, F.S., 2017. Dated phylogenetic studies of the southernmost American buthids (Scorpiones; Buthidae). *Mol. Phylogenet. Evol.* 110, 39–49.
- Olamendi-Portugal, T., Somodi, S., Fernández, J.A., Zamudio, F.Z., Becerril, B., Varga, Z., Panyi, G., Gáspár, R., Possani, L.D., 2005. Novel  $\alpha$ -KTx peptides from the venom of the scorpion *Centruroides elegans* selectively block Kv1.3 over IKCa1 K<sup>+</sup> channels of T cells. *Toxicon* 46, 418–429.
- Olamendi-Portugal, T., Bartok, A., Zamudio-Zuñiga, F., Balajthy, A., Becerril, B., Panyi, G., Possani, L.D., 2016. Isolation, chemical and functional characterization of several new K<sup>+</sup>-channel blocking peptides from the venom of the scorpion *Centruroides tecomanus*. *Toxicon* 115, 1–12.
- de Oliveira, U.C., Candido, D.M., Dorce, V.A.C., Junqueira-de Azevedo, I. de Loliola Meirelles, 2015. The transcriptome recipe for the venom cocktail of *Tityus bahiensis* scorpion. *Toxicon* 95, 52–61.
- de Oliveira, U.C., Nishiyama Jr., M.Y., dos Santos, M.B.V., de Paula Santos-da Silva, A., de Menezes Chalkidis, H., Souza-Imberg, A., Candido, D.M., Yamanouye, N., Dorce, V.A.C., Junqueira-de, I.D.L.M., et al., 2018. Proteomic endorsed transcriptomic profiles of venom glands from *Tityus obscurus* and *T. serrulatus* scorpions. *PLoS One* 13, e0193739.
- Ortiz, E., Gurrola, G.B., Schwartz, E.F., Possani, L.D., 2015. Scorpion venom components as potential candidates for drug development. *Toxicon* 93, 125–135.
- Otero, R., Navio, E., Céspedes, F., Núñez, M., Lozano, L., Moscoso, E., Matallana, C., Arsuza, N., García, J., Fernández, D., et al., 2004. Scorpion envenoming in two regions of Colombia: clinical, epidemiological and therapeutic aspects. *Trans. R. Soc. Trop. Med. Hyg.* 98, 742–750.
- Ozkan, O., Ciftci, G., 2010. Individual variation in the protein profile of the venom of *Mesobuthus gibbosus* (Brullé, 1832, Scorpiones: Buthidae) from Turkey. *J. Venom. Anim. Toxins Incl. Trop. Dis.* 16, 505–508.
- Ozkan, O., Kat, I., 2005. *Mesobuthus eupeus* scorpionism in Sanliurfa region of Turkey. *J. Venom. Anim. Toxins Incl. Trop. Dis.* 11, 479–491.
- Ozkan, O., Adigüzel, S., Yakişiran, S., Cesareti, Y., Orman, M., Karaer, K.Z., 2006. *Androctonus crassicauda* (Olivier 1807) scorpionism in the Sanliurfa Provinces of Turkey. *Head Neck* 3, 239–245.
- Ozkan, O., Uzun, R., Adigüzel, S., Cesareti, Y., Ertek, M., 2008. Evaluation of scorpion sting incidence in Turkey. *J. Venom. Anim. Toxins Incl. Trop. Dis.* 14, 128–140.
- Pardal, P.P., Ishikawa, E.A., Vieira, J.L., Coelho, J.S., Dórea, R.C., Abati, P.A., Quiroga, M.M., Chalkidis, H.M., 2014. Clinical aspects of envenomation caused by *Tityus obscurus* (Gervais, 1843) in two distinct regions of Pará state, Brazilian Amazon basin: a prospective case series. *J. Venom. Anim. Toxins Incl. Trop. Dis.* 20, 3.
- Parmakelis, A., Stathi, I., Chatzaki, M., Simaiakis, S., Spanos, L., Louis, C., Mylonas, M., 2006. Evolution of *Mesobuthus gibbosus* (Brullé, 1832)(scorpiones: Buthidae) in the northeastern mediterranean region. *Mol. Ecol.* 15, 2883–2894.
- Pipelzadeh, M.H., Jalali, A., Taraz, M., Pourabbas, R., Zaremirakabadi, A., 2007. An epidemiological and a clinical study on scorpionism by the Iranian scorpion *Hemiscorpius lepturus*. *Toxicon* 50, 984–992.
- Ponce Saavedra, J., Francke, O.F., 2004. Una nueva especie de alacrán del género *Centruroides* Marx (1890)(Scorpiones, Buthidae) de la depresión del Balsas, México. *Acta Zool. Mexic.* 20, 221–232.
- Porto, T.J., Brazil, T.K., Lira-da Silva, R.M., 2010. Scorpions, state of Bahia, northeastern Brazil. *Check List* 6, 292–297.
- Possani, L.D., Martin, B.M., Svendsen, I., 1982. The primary structure of noxiustoxin: a K<sup>+</sup>-channel blocking peptide, purified from the venom of the scorpion *Centruroides noxius* Hoffmann. *Carlsberg Res. Commun.* 47, 285–289.
- Possani, L.D., Becerril, B., Delepiere, M., Tytgat, J., 1999. Scorpion toxins specific for Na<sup>+</sup>-channels. *FEBS J.* 264, 287–300.
- Possani, L.D., Merino, E., Corona, M., Bolivar, F., Becerril, B., 2000. Peptides and genes coding for scorpion toxins that affect ion-channels. *Biochimie* 82, 861–868.
- Prendini, L., Esposito, L.A., 2010. A reanalysis of *Parabuthus* (Scorpiones: Buthidae) phylogeny with descriptions of two new *Parabuthus* species endemic to the Central Namib gravel plains, Namibia. *Zool. J. Linn. Soc.* 159, 673–710.
- Prendini, L., Wheeler, W.C., 2005. Scorpion higher phylogeny and classification, taxonomic anarchy, and standards for peer review in online publishing. *Cladistics* 21, 446–494.
- Pucca, M.B., Cerni, F.A., Junior, E.L.P., Bordon, K.d.C.F., Amorim, F.G., Cordeiro, F.A., Longhim, H.T., Cremonese, C.M., Oliveira, G.H., Arantes, E.C., 2015a. *Tityus serrulatus* venom—A lethal cocktail. *Toxicon* 108, 272–284.
- Pucca, M.B., Cerni, F.A., Peigneur, S., Bordon, K.C., Tytgat, J., Arantes, E.C., 2015b. Revealing the function and the structural model of Ts4: insights into the “non-toxic” toxin from *Tityus serrulatus* venom. *Toxins* 7, 2534–2550.
- Pucca, M.B., Peigneur, S., Cologna, C.T., Cerni, F.A., Zoccal, K.F., de CF Bordon, K., Faccioli, L.H., Tytgat, J., Arantes, E.C., 2015c. Electrophysiological characterization of the first *Tityus serrulatus* alpha-like toxin, Ts5: evidence of a pro-inflammatory toxin on macrophages. *Biochimie* 115, 8–16.
- Pucca, M.B., Bertolini, T.B., Cerni, F.A., Bordon, K.C., Peigneur, S., Tytgat, J., Bonato, V.L., Arantes, E.C., 2016a. Immunosuppressive evidence of *Tityus serrulatus* toxins Ts6 and Ts15: insights of a novel K<sup>+</sup> channel pattern in T cells. *Immunology* 147, 240–250.
- Pucca, M.B., Cerni, F.A., Cordeiro, F.A., Peigneur, S., Cunha, T.M., Tytgat, J., Arantes, E.C., 2016b. Ts8 scorpion toxin inhibits the Kv4.2 channel and produces nociception in vivo. *Toxicon* 119, 244–252.
- Quintero-Hernández, V., Jiménez-Vargas, J.M., Gurrola, G.B., Valdivia, H.H., Possani, L.D., 2013. Scorpion venom components that affect ion-channels function. *Toxicon* 76, 328–342.
- Quintero-Hernández, V., Ramírez-Carreto, S., Romero-Gutiérrez, M.T., Valdez-Velázquez, L.L., Becerril, B., Possani, L.D., Ortiz, E., 2015. Transcriptomic analysis of scorpion species belonging to the *Vaejovis* genus. *PLoS One* 10, e0117188.
- Radmanesh, M., 1990. *Androctonus crassicauda* sting and its clinical study in Iran. *J. Trop. Med. Hyg.* 93, 323–326.
- Ramírez, A.N., Gurrola, G.B., Martin, B.M., Possani, L.D., 1988. Isolation of several toxins from the venom of the scorpion *Centruroides limpidus tecomanus* Hoffmann. *Toxicon* 26, 773–783.
- Ramírez-Carreto, S., Quintero-Hernández, V., Jiménez-Vargas, J.M., Corzo, G., Possani, L.D., Becerril, B., Ortiz, E., 2012. Gene cloning and functional characterization of four novel antimicrobial-like peptides from scorpions of the family Vaejovidae. *Peptides* 34, 290–295.
- Razi, E., Malekanrad, E., 2008. Asymmetric pulmonary edema after scorpion sting: a case report. *Rev. do Inst. Med. Trop. São Paulo* 50, 347–350.
- Rein, J.O., 2018. **The Scorpion Files.** <https://www.ntnu.no/ub/scorpion-files>.
- Rendón-Anaya, M., Delaye, L., Possani, L.D., Herrera-Estrella, A., 2012. Global transcriptome analysis of the scorpion *Centruroides noxius*: new toxin families and evolutionary insights from an ancestral scorpion species. *PLOS One* 7, e43331.
- Restano-Cassulini, R., Olamendi-Portugal, T., Zamudio, F., Becerril, B., Possani, L.D., 2008. Two novel ergotoxins, blockers of K<sup>+</sup>-channels, purified from the Mexican scorpion *Centruroides elegans elegans*. *Neurochem. Res.* 33, 1525–1533.
- Riño-Umbarila, L., Rodríguez-Rodríguez, E.R., Santibáñez-López, C.E., Güiereca, L., Uribe-Romero, S.J., Gómez-Ramírez, I.V., Cárcamo-Noriega, E.N., Possani, L.D., Becerril, B., 2017. Updating knowledge on new medically important scorpion species in Mexico. *Toxicon* 138, 130–137.
- Rokytka, D.R., Ward, M.J., 2017. Venom-gland transcriptomics and venom proteomics of the blackback scorpion (*Hadrurus spadix*) reveal detectability challenges and an unexplored realm of animal toxin diversity. *Toxicon* 128, 23–37.
- de Roodt, A.R., 2014. Comments on environmental and sanitary aspects of the scorpionism by *Tityus trivittatus* in Buenos Aires City, Argentina. *Toxins* 6, 1434–1452.
- de Roodt, A.R., García, S.I., Salomón, O.D., Segre, L., Dolab, J.A., Funes, R.F., de Titto, E.H., 2003. Epidemiological and clinical aspects of scorpionism by *Tityus trivittatus* in Argentina. *Toxicon* 41, 971–977.
- de Roodt, A.R., Lago, N.R., Salomón, O.D., Laskowicz, R.D., de Román, L.E.N., López, R.A., Montero, T.E., Vega, V. d. V., 2009. A new venomous scorpion responsible for severe envenomation in Argentina: *Tityus confluentis*. *Toxicon* 53, 1–8.
- Rosin, R., Shulov, A., 1963. Studies on the scorpion *Nebo hieronchonticus*. *J. Zool.* 140, 547–575.
- Saad, K., El-Hamed, M.A.A., Abo-Elela, M.G.M., Ahmed, A.E., Abdel-Baseer, K.A., Aboul-Khair, M.D., Metwally, K.A., El-Houfey, A.A., Hasan, G.M., El-Shareef, A.M., 2017. Neurologic complications in children with scorpionism: a retrospective study in upper Egypt. *J. Child Neurol.* 32, 537–542.
- Salazar, M.H., Arenas, I., Corrales-García, L.L., Miranda, R., Vélez, S., Sánchez, J., Mendoza, K., Cleghorn, J., Zamudio, F.Z., Castillo, A., et al., 2018. Venoms of *Centruroides* and *Tityus* species from Panama and their main toxic fractions. *Toxicon* 141, 79–87.
- Salem, M.L., Shoukry, N.M., Teleb, W.K., Abdel-Daim, M.M., Abdel-Rahman, M.A., 2016. In vitro and in vivo antitumor effects of the Egyptian scorpion *Androctonus amoreuxi* venom in an Ehrlich ascites tumor model. *SpringerPlus* 5, 570.
- Salman, M.M., Hammad, S., 2017. Oxidative stress and some biochemical alterations due to scorpion (*Leiurus quinquestriatus*) crude venom in rats. *Biomed. Pharmacother.* 91, 1017–1021.
- Sanaei-Zadeh, H., Marashi, S.M., Dehghani, R., 2017. Epidemiological and clinical characteristics of scorpionism in Shiraz (2012–2016); development of a clinical severity grading for Iranian scorpion envenomation. *Med. J. Islam. Repub. Iran MJIRI* 31, 158–166.
- Santibáñez-López, C.E., Francke, O.F., Ureta, C., Possani, L.D., 2015. Scorpions from Mexico: from species diversity to venom complexity. *Toxins* 8. <https://doi.org/10.3390/toxins8010002>.
- Santibáñez-López, C.E., Cid-Urbe, J.I., Batista, C.V., Ortiz, E., Possani, L.D., 2016. Venom gland transcriptomic and proteomic analyses of the enigmatic scorpion *Superstitionia donensis* (Scorpiones: Superstitioniidae), with insights on the evolution of its venom components. *Toxins* 8, 367.
- Santibáñez-López, C.E., Cid-Urbe, J.I., Zamudio, F.Z., Batista, C.V., Ortiz, E., Possani, L.D., 2017. Venom gland transcriptomic and venom proteomic analyses of the scorpion *Megacormus gertschi* Díaz-Najera, 1966 (Scorpiones: Euscorpidae: Megacorminae). *Toxicon* 133, 95–109.
- Santos, M.S., Silva, C.G., Neto, B.S., Júnior, C.R.G., Lopes, V.H., Júnior, A.G.T., Bezerra, D.A., Luna, J.V., Cordeiro, J.B., Júnior, J.G., et al., 2016. Clinical and epidemiological aspects of scorpionism in the world: a systematic review. *Wilderness Environ. Med.* 27, 504–518.
- Sari, A., Hosseini, S., et al., 2011. History of study and checklist of the scorpion fauna (Arachnida: scorpiones) of Iran. *Prog. Biol. Sci.* 1, 16–23.
- Schmitt, C., Torrents, R., Simon, N., de Haro, L., 2017. First described envenomation by *Centruroides pockockii* scorpion in the French caribbean island Guadeloupe. *Wilderness Environ. Med.* 28, 159–160.
- Schwartz, E.F., Diego-García, E., de la Vega, R.C.R., Possani, L.D., 2007. Transcriptome analysis of the venom gland of the Mexican scorpion *Hadrurus gertschi* (Arachnida: scorpiones). *BMC Genom.* 8, 119.
- Seiter, M., Koç, H., Ullrich, A., Yağmur, E.A., 2016. The case history of a toxic sting of a *Leiurus abdullahbayrami* scorpion in Turkey. *Arachnol. Mittl./Arachnol. Lett.* 51, 64–66.
- Serrano, S.M., 2013. The long road of research on snake venom serine proteinases. *Toxicon* 62, 19–26.
- Shahi, M., Rafinejad, J., Az-Khosravi, L., Moosavy, S.H., 2015. First report of death due to *Hemiscorpius acanthocercus* envenomation in Iran: case report. *Electron. Physician* 7, 1234.
- Shahi, M., Moosavy, S.H., Rafinejad, J., Zare, S., Navidpour, S., Madani, A., 2016. Epidemiological and clinical aspects of scorpion sting among children in south part of

- Iran. Global J. Health Sci. 9, 289.
- Sharma, P.P., Fernández, R., Esposito, L.A., González-Santillán, E., Monod, L., 2015. Phylogenomic resolution of scorpions reveals multilevel discordance with morphological phylogenetic signal. *Proc. R. Soc. Lond. B Biol. Sci.* 282, 20142953.
- Silva, B. A. J. d., Fé, N.F., Gomes, A. A. d. S., Souza, A.d.S., Sachtet, J.d.A.G., Fan, H.W., Melo, G.C.d., Monteiro, W.M., 2017. Implication of *Tityus apiacas* (Loureiro, 2002) in scorpion envenomations in the southern amazon border, Brazil. *Rev. Soc. Bras. Med. Trop.* 50, 427–430.
- Sissom, W.D., Lourenço, W.R., 1987. The genus *Centruroides* in South America (scorpiones, Buthidae). *J. Arachnol.* 11–28.
- Skolnik, A.B., Ewald, M.B., 2013. Pediatric scorpion envenomation in the United States: morbidity, mortality, and therapeutic innovations. *Pediatr. Emerg. Care* 29, 98–103.
- Sofer, S., Gueron, M., 1988. Respiratory failure in children following envenomation by the scorpion *Leiurus quinquestriatus*: hemodynamic and neurological aspects. *Toxicon* 26, 931–939.
- Sofer, S., Shalev, H., Weizman, Z., Shahak, E., Gueron, M., 1991. Acute pancreatitis in children following envenomation by the yellow scorpion *Leiurus quinquestriatus*. *Toxicon* 29, 125–128.
- De Sousa, L., Boadas, J., Kiriakos, D., Borges, A., Boadas, J., Marcano, J., Turkali, I., De Los Ríos, M., 2007. Scorpionism due to *Tityus neoespartanus* (scorpiones, Buthidae) in margarita island, northeastern Venezuela. *Rev. Soc. Bras. Med. Trop.* 40, 681–685.
- Sousa, P., Arnedo, M.A., Harris, D.J., 2017. Updated catalogue and taxonomic notes on the old-world scorpion genus *Buthus* Leach, 1815 (scorpiones, Buthidae). *ZooKeys* 686, 15–84.
- Strong, P., Clark, G., Armugam, A., De-Allie, F., Joseph, J., Yemul, V., Deshpande, J., Kamat, R., Gadre, S., Gopalakrishnakone, P., et al., 2001. Tamulustoxin: a novel potassium channel blocker from the venom of the Indian red scorpion *Mesobuthus tamulus*. *Arch. Biochem. Biophys.* 385, 138–144.
- Strong, P.N., Mukherjee, S., Shah, N., Chowdhary, A., Jeyaseelan, K., 2015. Scorpion venom research around the world: Indian Red Scorpion. In: *Scorpion Venoms*. Springer, pp. 369–382.
- Stuber, M., Nentwig, W., 2016. How informative are case studies of spider bites in the medical literature? *Toxicon* 114, 40–44.
- Suresh, S.S., Zaki, H., Shalamzari, J.E., Bhatnagar, G., 2014. Osteomyelitis calcaneum due to a scorpion sting. *J. Foot Ankle Surg.* 53, 340–343.
- Teixeira Jr., A., Fontoura, B., Freire-Maia, L., Machado, C., Camargos, E., Teixeira, M., 2001. Evidence for a direct action of *Tityus serrulatus* scorpion venom on the cardiac muscle. *Toxicon* 39, 703–709.
- Teruel, R., 2008. Confirmation of the occurrence of *Centruroides gracilis* (Latreille 1805) (scorpiones: Buthidae) in Jamaica. *Bol. Soc. Entomol. Aragon.* 42, 370.
- Teruel, R., de los Santos, G., 2018. Two new *Tityus* CL Koch, 1836 (scorpiones: Buthidae) from hispaniola, greater antilles. *Euscorpius* 2018, 1–16.
- Teruel, R., Ponce-Saavedra, J., Quijano-Ravell, A.F., 2015. Redescription of *Centruroides noxius* and description of a closely related new species from western Mexico (Scorpiones: Buthidae). *Rev. Mex. Biodivers.* 86, 896–911.
- Torabi, E., Behdani, M., Chafi, M.H., Moazzami, R., Sabatier, J.-M., Khalaj, V., Shahbazzadeh, D., Bagheri, K.P., 2017. Characteristics and lethality of a novel recombinant dermonecrotic venom phospholipase D from *Hemiscorpius lepturus*. *Toxins* 9, 102.
- Torrez, P.P., Quiroga, M.M., Abati, P.A., Mascheretti, M., Costa, W.S., Campos, L.P., França, F.O., 2015. Acute cerebellar dysfunction with neuromuscular manifestations after scorpionism presumably caused by *Tityus obscurus* in Santarém, Pará/Brazil. *Toxicon* 96, 68–73.
- Touloun, O., Slimani, T., Boumezzough, A., 2001. Epidemiological survey of scorpion envenomation in southwestern Morocco. *J. Venom. Anim. Toxins* 7, 199–218.
- Touloun, O., Boumezzough, A., Slimani, T., 2012. Scorpion envenomation in the region of Marrakesh Tensift Alhaouz (Morocco): epidemiological characterization and therapeutic approaches. *Serket* 13, 38–50.
- Trejo, E., Borges, A., Nañez, B., de Becemborg, I.L., de Alfonso, R.G., Alfonso, M.J., 2012. *Tityus zuliaanus* venom induces massive catecholamine release from PC12 cells and in a mouse envenomation model. *Toxicon* 59, 117–123.
- Undheim, E.A., Jones, A., Clauser, K.R., Holland, J.W., Pineda, S.S., King, G.F., Fry, B.G., 2014. Clawing through evolution: toxin diversification and convergence in the ancient lineage Chilopoda (Centipedes). *Mol. Biol. Evol.* 31, 2124–2148.
- Undheim, E.A., Hamilton, B.R., Kurniawan, N.D., Bowlay, G., Cribb, B.W., Merritt, D.J., Fry, B.G., King, G.F., Venter, D.J., 2015. Production and packaging of a biological arsenal: evolution of centipede venoms under morphological constraint. *Proc. Natl. Acad. Sci. U. S. A.* 112, 4026–4031.
- Valdez-Cruz, N.A., Dávila, S., Licea, A., Corona, M., Zamudio, F.Z., García-Valdes, J., Boyer, L., Possani, L.D., 2004. Biochemical, genetic and physiological characterization of venom components from two species of scorpions: *Centruroides exilicauda* Wood and *Centruroides sculpturatus* Ewing. *Biochimie* 86, 387–396.
- Valdez-Velazquez, L.L., Quintero-Hernández, V., Romero-Gutiérrez, M.T., Coronas, F.I.V., Possani, L.D., 2013. Mass fingerprinting of the venom and transcriptome of venom gland of scorpion *Centruroides tecomanus*. *PLoS One* 8, e66486.
- Valdez-Velazquez, L., Romero-Gutierrez, M., Delgado-Enciso, I., Dobrovinskaya, O., Melnikov, V., Quintero-Hernández, V., Ceballos-Magaña, S., Gaitan-Hinojosa, M., Coronas, F., Puebla-Perez, A., et al., 2016. Comprehensive analysis of venom from the scorpion *Centruroides tecomanus* reveals compounds with antimicrobial, cytotoxic, and insecticidal activities. *Toxicon* 118, 95–103.
- Vandendriessche, T., Olamendi-Portugal, T., Zamudio, F.Z., Possani, L.D., Tytgat, J., 2010. Isolation and characterization of two novel scorpion toxins: the  $\alpha$ -toxin-like Cell8, specific for Nav1.7 channels and the classical anti-mammalian Cell9, specific for Nav1.4 channels. *Toxicon* 56, 613–623.
- de la Vega, R.C.R., Possani, L.D., 2004. Current views on scorpion toxins specific for  $K^+$  channels. *Toxicon* 43, 865–875.
- de la Vega, R.C.R., Schwartz, E.F., Possani, L.D., 2010. Mining on scorpion venom biodiversity. *Toxicon* 56, 1155–1161.
- Veisesh, M., Gabikian, P., Bahrami, S.-B., Veisesh, O., Zhang, M., Hackman, R.C., Ravanpay, A.C., Stroud, M.R., Kusuma, Y., Hansen, S.J., et al., 2007. Tumor paint: a chlorotoxin: cy5.5 bioconjugate for intraoperative visualization of cancer foci. *Cancer Res.* 67, 6882–6888.
- Wang, G., Strichartz, G., 1983. Purification and physiological characterization of neurotoxins from venoms of the scorpions *Centruroides sculpturatus* and *Leiurus quinquestriatus*. *Mol. Pharmacol.* 23, 519–533.
- Ward, M.J., Ellsworth, S.A., Rokyta, D.R., 2018. Venom-gland transcriptomics and venom proteomics of the Hentz striped scorpion (*Centruroides hentzi*; Buthidae) reveal high toxin diversity in a harmless member of a lethal family. *Toxicon* 142, 14–29.
- Whittington, A.C., Mason, A.J., Rokyta, D.R., 2018. A single mutation unlocks cascading exaptations in the origin of a potent pitviper neurotoxin. *Mol. Biol. Evol.* 35, 887–898.
- Wickham, H., 2016. *ggplot2: Elegant Graphics for Data Analysis*. Springer.
- Yağmur, E.A., Yalçın, M., Çalısır, G., 2008. Distribution of *Androctonus crassicauda* (Olivier 1807) and *Buthacus macrocentrus* (Ehrenberg 1828)(scorpiones: Buthidae) in Turkey. *Serket* 11, 13–18.
- Zamudio, F., Saavedra, R., Martin, B.M., Gurrola-Briones, G., Héron, P., Possani, L.D., 1992. Amino acid sequence and immunological characterization with monoclonal antibodies of two toxins from the venom of the scorpion *Centruroides noxius* Hoffmann. *FEBS J.* 204, 281–292.
- Zeng, X.-C., Corzo, G., Hahin, R., 2005. Scorpion venom peptides without disulfide bridges. *IUBMB Life* 57, 13–21.
- Zhang, L., Shi, W., Zeng, X.-C., Ge, F., Yang, M., Nie, Y., Bao, A., Wu, S., Guojij, E., 2015. Unique diversity of the venom peptides from the scorpion *Androctonus bicolor* revealed by transcriptomic and proteomic analysis. *J. Proteom.* 128, 231–250.
- Zoccal, K.F., da Silva Bitencourt, C., Secatto, A., Sorgi, C.A., Bordon, K.d.C.F., Sampaio, S.V., Arantes, E.C., Faccioli, L.H., 2011. *Tityus serrulatus* venom and toxins Ts1, Ts2 and Ts6 induce macrophage activation and production of immune mediators. *Toxicon* 57, 1101–1108.
- Zoccal, K.F., da Silva Bitencourt, C., Sorgi, C.A., Bordon, K.d.C.F., Sampaio, S.V., Arantes, E.C., Faccioli, L.H., 2013. Ts6 and Ts2 from *Tityus serrulatus* venom induce inflammation by mechanisms dependent on lipid mediators and cytokine production. *Toxicon* 61, 1–10.